

# Package ‘CytoMethIC’

September 11, 2025

**Type** Package

**Title** DNA methylation-based machine learning models

**Description** This package provides model data and functions for easily using machine learning models that use data from the DNA methylome to classify cancer type and phenotype from a sample. The primary motivation for the development of this package is to abstract away the granular and accessibility-limiting code required to utilize machine learning models in R. Our package provides this abstraction for RandomForest, e1071 Support Vector, Extreme Gradient Boosting, and Tensorflow models. This is paired with an ExperimentHub component, which contains models developed for epigenetic cancer classification and predicting phenotypes. This includes CNS tumor classification, Pan-cancer classification, race prediction, cell of origin classification, and subtype classification models. The package links to our models on ExperimentHub. The package currently supports HM450, EPIC, EPICv2, MSA, and MM285.

**Version** 1.5.0

**License** Artistic-2.0

**Depends** R (>= 4.4.0), ExperimentHub

**Imports** utils, stats, tools, sesame, methods, sesameData,  
BiocParallel, BiocManager

**VignetteBuilder** knitr

**Suggests** tibble, BiocStyle, randomForest, testthat, knitr, rmarkdown,  
e1071, xgboost, keras, tensorflow

**URL** <https://github.com/zhou-lab/CytoMethIC>

**BugReports** <https://github.com/zhou-lab/CytoMethIC/issues>

**biocViews** ExperimentData, MicroarrayData, Genome, ExperimentHub,  
MethylationArrayData, CancerData, PackageTypeData

**NeedsCompilation** no

**RoxygenNote** 7.3.2

**Encoding** UTF-8

**git\_url** <https://git.bioconductor.org/packages/CytoMethIC>

**git\_branch** devel

**git\_last\_commit** 88f8336

**git\_last\_commit\_date** 2025-04-15

**Repository** Bioconductor 3.22

**Date/Publication** 2025-09-11

**Author** Wanding Zhou [aut] (ORCID: <<https://orcid.org/0000-0001-9126-1932>>),  
Jacob Fanale [aut, cre] (ORCID:  
<<https://orcid.org/0009-0002-0490-3269>>)

**Maintainer** Jacob Fanale <jfanale@seas.upenn.edu>

## Contents

.optimizeFrac . . . . .	2
cmi_checkVersion . . . . .	3
cmi_deconvolution . . . . .	3
cmi_deconvolution2 . . . . .	4
cmi_models . . . . .	5
cmi_predict . . . . .	5

<b>Index</b>	<b>7</b>
--------------	----------

---

.optimizeFrac	<i>Internal function for fraction optimization</i>
---------------	--

---

## Description

Internal function for fraction optimization

## Usage

```
.optimizeFrac(
  frac,
  ref,
  q,
  errFunc,
  temp = 0.5,
  maxIter = 1000,
  delta = 1e-04,
  step.max = 1,
  verbose = FALSE
)
```

## Arguments

frac	initial fraction
ref	reference
q	query
errFunc	error function
temp	annealing temperature
maxIter	maximum iteration to stop after converge
delta	delta score to reset counter
step.max	maximum step, do not adjust
verbose	output debug info

**Value**

a list of fractions and min err

---

cmi_checkVersion	<i>Check CytoMethIC versions</i>
------------------	----------------------------------

---

**Description**

print package version of cytomethic and depended packages to help troubleshoot installation issues.

**Usage**

```
cmi_checkVersion()
```

**Value**

print the versions of cytomethic and dependencies

**Examples**

```
cmi_checkVersion()
```

---

cmi_deconvolution	<i>Reference-based cell type deconvolution</i>
-------------------	--

---

**Description**

This is a reference-based cell composition estimation. The function takes a reference methylation status matrix (rows for probes and columns for cell types) and a query beta value measurement.

**Usage**

```
cmi_deconvolution(ref, q, trim = FALSE, ...)
```

**Arguments**

ref	reference methylation
q	target measurement: length(q) == nrow(ref)
trim	to trim query input beta values. this relieves unclean background subtraction
...	extra parameters for optimization.

**Details**

The length of the target beta values should be the same as the number of rows of the reference Matrix. The function outputs a list containing the estimated cell fraction, the error of optimization.

**Value**

a list of fraction, min error.

**Examples**

```
ref = cbind(
  CD4 = c(1,1,1,0,1,0),
  CD19 = c(0,0,1,1,0,1),
  CD14 = c(1,1,1,1,0,1))
rownames(ref) = paste0("cg",1:6)
trueFrac = runif(3)
trueFrac = trueFrac / sum(trueFrac)
q = ref %*% trueFrac
trueFrac
cmi_deconvolution(ref, q)
```

---

cmi_deconvolution2	<i>Reference-based cell type deconvolution (allowing one unknown component)</i>
--------------------	---

---

**Description**

This is a reference-based cell composition estimation. The function takes a reference methylation status matrix (rows for probes and columns for cell types) and a query beta value measurement.

**Usage**

```
cmi_deconvolution2(ref, q, trim = FALSE, ...)
```

**Arguments**

ref	reference methylation
q	target measurement: length(q) == nrow(ref)
trim	to trim query input beta values. this relieves unclean background subtraction
...	extra parameters to .optimizeFrac

**Details**

The length of the target beta values should be the same as the number of rows of the reference Matrix. The method assumes one unknown component. It outputs a list containing the estimated cell fraction, the error of optimization and methylation status of the unknown component.

**Value**

a list of fraction, min error and unknown component methylation state

**Examples**

```
ref = cbind(
  CD4 = c(1,1,1,0,1,0),
  CD19 = c(0,0,1,1,0,1),
  CD14 = c(1,1,1,1,0,1))
rownames(ref) = paste0("cg",1:6)
trueFrac = runif(4)
trueFrac = trueFrac / sum(trueFrac)
```

```

ref_unk = sample(c(0,1), nrow(ref), replace=TRUE)
q = cbind(ref_unk, ref) %% trueFrac
trueFrac
res = cmi_deconvolution2(ref, q)
res$frac

```

---

cmi\_models

*Master data frame for all model objects*


---

### Description

This is an internal object which will be updated on every new release

### Format

tibble

### Value

master sheet of CytoMethIC model objects

### Examples

```
print(cmi_models[,c("EHID", "Title")])
```

---

cmi\_predict

*The cmi\_predict function takes in a model and a sample, and uses the model to predict it. This function supports randomForest, e1071::svm, xgboost, and keras/tensorflow models. For xgboost and keras models, the features used in classification as well as a label mapping must be provided for output.*

---

### Description

The cmi\_predict function takes in a model and a sample, and uses the model to predict it. This function supports randomForest, e1071::svm, xgboost, and keras/tensorflow models. For xgboost and keras models, the features used in classification as well as a label mapping must be provided for output.

### Usage

```
cmi_predict(betas, cmi_model, verbose = FALSE, BPPARAM = SerialParam())
```

### Arguments

betas	DNA methylation beta
cmi_model	Cytomethic model downloaded from ExperimentHub
verbose	be verbose with warning
BPPARAM	use MulticoreParam(n) for parallel processing

**Value**

predicted cancer type label

**Examples**

```
library(sesame)
library(ExperimentHub)
library(CytoMethIC)

## Cancer Type
model = ExperimentHub()[["EH8395"]]
betas = openSesame(sesameDataGet("EPICv2.8.SigDF")[[1]])
betas = imputeBetas(mLiftOver(betas, "HM450"))
cmi_predict(betas, model)

betas = openSesame(sesameDataGet('EPIC.1.SigDF'), mask=FALSE)
cmi_predict(betas, model)

betas = sesameDataGet("HM450.1.TCGA.PAAD")$betas
betas = imputeBetas(betas)
cmi_predict(betas, model)
```

# Index

.optimizeFrac, 2  
cmi\_checkVersion, 3  
cmi\_deconvolution, 3  
cmi\_deconvolution2, 4  
cmi\_models, 5  
cmi\_predict, 5