

# Package ‘cfdnakit’

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**Title** Fragment-length analysis package from high-throughput sequencing of cell-free DNA (cfDNA)

**Version** 1.4.0

**Description** This package provides basic functions for analyzing shallow whole-genome sequencing (~0.3X or more) of cell-free DNA (cfDNA). The package basically extracts the length of cfDNA fragments and aids the visualization of fragment-length information. The package also extract fragment-length information per non-overlapping fixed-sized bins and used it for calculating ctDNA estimation score (CES).

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## Contents

cfdnakit-package	2
calculate_CES_score	4
call_cnv	5
create_blacklist_gr	6
create_PoN	6
extract_insert_size	7
filter_read_on_blacklist	7
fragment_dist	8
get_fragment_profile	8
get_segment_byresolution	9
get_solution_table	10
get_zscore_profile	10
GRCh2UCSCGRanges	11
if_exist_baifile	12
if_ucsc_chrformat	12
make_density_table	13
overlap_bin_with_segment	13
plot_cnv_solution	14
plot_distance_matrix	15
plot_fragment_dist	15
plot_sl_ratio	16
plot_transformed_sl	17
read_bamfile	18
read_PoN_files	19
segmentByPSCB	19
test_ysize_KolmogorovSmirnov	20
UCSC2GRChSampleBam	20
util.bias_correct	21
zscore_transform	21
%>%	22

<b>Index</b>	<b>23</b>
--------------	-----------

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cfdnakit-package	<i>Fragmen-length analysis package from high-throughput sequencing of cell-free DNA (cfDNA)</i>
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## Description

This package provides basic functions for analyzing shallow whole-genome sequencing (~0.3X or more) of cell-free DNA (cfDNA). The package basically extracts the length of cfDNA fragments and aids the visualization of fragment-length information. The package also extract fragment-length information per non-overlapping fixed-sized bins and used it for calculating ctDNA estimation score (CES).

## Details

This package provides functions for analyzing using shallow whole-genome sequencing data (~0.3X or more) of circulating cell-free DNA (cfDNA). The aim is to estimate circulating tumor DNA using its characteristic short-fragmented cfDNA. The package extracts length of each cfDNA and assists the visualization of fragment-length distribution. A short-fragment ratio is calculated per non-overlapping fixed-sized bins. Genome-wide copy-number alteration is estimated by the short-fragmented cfDNA. The ctDNA estimation score (CES) comprehensively estimates the circulating tumor DNA based on the short-fragment analysis.

## Author(s)

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## Examples

```
library(cfdnakit)
## Reading in a bamfile
sample_bamfile = system.file("extdata",
                             "ex.plasma.bam",
                             package = "cfdnakit")
plasma_SampleBam = read_bamfile(sample_bamfile,
                               apply_blacklist = FALSE)

## Plot a fragment-length distribution of a sample
plot_fragment_dist(list("Plasma.Sample"=plasma_SampleBam))

## Plot a fragment-length distribution of two samples
control_RDS_file =
  system.file("extdata", "BH01_CHR15.SampleBam.rds",
             package = "cfdnakit")
### Load example SampleBam of Healthy cfDNA
control_bins =
  readRDS(control_RDS_file)

comparing_list = list("Healthy.cfDNA"=control_bins,
                     "Patient.1"=plasma_SampleBam)
plot_fragment_dist(comparing_list)

## Derived and plot genome-wide short-fragment cfDNA
patient.SampleFragment =
  get_fragment_profile(plasma_SampleBam,
                      sample_id = "Patient.1")
plot_sl_ratio(patient.SampleFragment)

## Derived and plot normalized short-fragment cfDNA
PoN_rdsfile = system.file(
  "extdata",
  "ex.PoN.rds",
  package = "cfdnakit")
## Loading example PoN data
PoN.profiles = readRDS(PoN_rdsfile)

sample_zscore =
  get_zscore_profile(patient.SampleFragment,
```

```
      PoN.profiles)
sample_zscore_segment = segmentByPSCB(sample_zscore)
plot_transformed_sl(sample_zscore, sample_zscore_segment)

## Estimate circulating tumor DNA
calculate_CES_score(sample_zscore_segment)
```

---

calculate\_CES\_score     *Calculate CES Score from Segmentation*

---

## Description

Calculate CES Score from Segmentation

## Usage

```
calculate_CES_score(sample_segmentation)
```

## Arguments

```
sample_segmentation
  Segmentation Dataframe
```

## Value

Numeric; CES score

## Examples

```
### Loading example SampleBam file
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bambin <- readRDS(example_file)
### Example PoN
PoN_rdsfile <- system.file("extdata", "ex.PoN.rds", package = "cfdnakit")
pon_profiles <- readRDS(PoN_rdsfile)
sample_profile <- get_fragment_profile(sample_bambin, sample_id = "Patient1")

sample_zscore <- get_zscore_profile(sample_profile, pon_profiles)

sample_zscore_segment <- segmentByPSCB(sample_zscore)

calculate_CES_score(sample_zscore_segment)
```

---

`call_cnv`*Call Copy-number Variation from SLRatio and segmentation*

---

**Description**

Call Copy-number Variation from SLRatio and segmentation

**Usage**

```
call_cnv(  
  sample_segmentation,  
  sample_zscore,  
  callChr = seq_len(22),  
  tfs = c(0, 0.7),  
  ploidies = c(1.5, 3),  
  MaxCN = 4  
)
```

**Arguments**

<code>sample_segmentation</code>	segmentation dataframe from <code>segmentByPSCBS</code>
<code>sample_zscore</code>	zscore dataframe
<code>callChr</code>	chromosome to analysis : Default <code>c(1:22)</code>
<code>tfs</code>	range of fitting tumor fraction : Default <code>c(0,0.8)</code>
<code>ploidies</code>	range of fitting chromosomal ploidy : Default <code>c(1.5,4)</code>
<code>MaxCN</code>	maximum copy-number : Default 4

**Value**

List of cnvcalling solutions

**Examples**

```
### Loading example SampleBam file  
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")  
sample_bambin <- readRDS(example_file)  
### Example PoN  
PoN_rdsfile <- system.file("extdata", "ex.PoN.rds", package = "cfdnakit")  
pon_profiles <- readRDS(PoN_rdsfile)  
sample_profile <- get_fragment_profile(sample_bambin, sample_id = "Patient1")  
  
sample_zscore <- get_zscore_profile(sample_profile, pon_profiles)  
  
sample_zscore_segment <- segmentByPSCB(sample_zscore)  
  
sample_cnv <- call_cnv(sample_zscore_segment, sample_zscore, tfs=c(0.1,0.3), ploidies=c(1.5,2), MaxCN=3)  
plot_cnv_solution(sample_cnv, selected_solution = 1)
```

---

create\_blacklist\_gr     *Create Blacklist regions GRanges object*

---

**Description**

Create Blacklist regions GRanges object

**Usage**

```
create_blacklist_gr(blacklist_files)
```

**Arguments**

blacklist\_files  
Character; Filepath to file containing blacklist regions

**Value**

GRanges object of blacklist regions

---

create\_PoN                 *Create Panel-of-Normal (PoN) object*

---

**Description**

Create Panel-of-Normal (PoN) object

**Usage**

```
create_PoN(list_rdsfiles)
```

**Arguments**

list\_rdsfiles     Character; a file contains paths to Profile.Rdata per line

**Value**

Null

**Examples**

```
healthy.1 <- system.file("extdata", "ex.healthy1.rds", package = "cfdnakit")
healthy.2 <- system.file("extdata", "ex.healthy2.rds", package = "cfdnakit")

path_to_PoN_txt <- paste0(system.file("extdata", package = "cfdnakit"), "/temp.reference_healthy.listfile")
fileConn<-file(path_to_PoN_txt)
writelines(c(healthy.1, healthy.2), fileConn)
close(fileConn)

PoN.profiles <- create_PoN(path_to_PoN_txt)
file.remove(path_to_PoN_txt)
```

---

extract\_insert\_size     *Extract Insert size from SampleBam*

---

**Description**

Extract Insert size from SampleBam

**Usage**

```
extract_insert_size(readbam_bin, maximum_length = 600, minimum_length = 20)
```

**Arguments**

readbam\_bin     SampleBam Object

maximum\_length   Int; Maximum length of fragment. cfDNA fragment longer than this value will not be considered; Default 600

minimum\_length   Int; Minimum length of fragment. cfDNA fragment shorter than this value will not be considered; Default 20

**Value**

Numeric Vector; Insert size of given sample

**Examples**

```
### Loading example SampleBam file
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bam_bin <- readRDS(example_file)
extract_insert_size(sample_bam_bin)
### Extract only insert size of fragment having specific size
extract_insert_size(sample_bam_bin, maximum_length=500, minimum_length = 50)
```

---

filter\_read\_on\_blacklist

*Filter out reads on blacklist regions*

---

**Description**

Filter out reads on blacklist regions

**Usage**

```
filter_read_on_blacklist(sample_bin, blacklist_files = NULL, genome = "hg19")
```

**Arguments**

sample\_bin     SampleBam; Object from function read\_bamfile

blacklist\_files     Character; Filepath to file containing blacklist regions

genome     Character; Abbreviation of reference genome; Either hg19 or mm10. default:hg19

**Value**

SampleBam after filtering out read on balck list regions

---

fragment_dist	<i>Get insert-size distribution table</i>
---------------	---

---

**Description**

Get insert-size distribution table

**Usage**

```
fragment_dist(readbam_bin, maximum_length = 600, minimum_length = 20)
```

**Arguments**

readbam_bin	SampleBam Object from function read_bamfile
maximum_length	Int; Maximum length of fragment. cfDNA fragment longer than this value will not be considered; Default 600
minimum_length	Int; Minimum length of fragment. cfDNA fragment shorter than this value will not be considered; Default 20

**Value**

Distribution table of fragment length

---

get_fragment_profile	<i>Getting fragment-length information</i>
----------------------	--

---

**Description**

Getting fragment-length information

**Usage**

```
get_fragment_profile(
  readbam_bin,
  sample_id,
  genome = "hg19",
  short_range = c(100, 150),
  long_range = c(151, 250),
  maximum_length = 600,
  minimum_length = 20
)
```



**Arguments**

readbam_bin	SampleBam Object
sample_id	Character; Given sample ID
genome	abbreviation of reference genome; namely hg19, mm10. default:hg19
short_range	Vector of 2 Int; Range of fragment length to be defined as short fragment; Default c(100,150)
long_range	Vector of 2 Int; Range of fragment length to be defined as long fragment; Default c(151,250)
maximum_length	Int; Maximum length of fragment. cfDNA fragment longer than this value will not be considered; Default 600
minimum_length	Int; Minimum length of fragment. cfDNA fragment shorter than this value will not be considered; Default 20

**Value**

SampleFragment Object; Fragment length information for quality check and downstream analysis per bin and summary of sample

**Examples**

```
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bam_bin <- readRDS(example_file)
sample_profile <- get_fragment_profile(sample_bam_bin, sample_id = "Patient1")
```

---

get\_segment\_bysolution

*Return CNV segmentation result from given all CNV solutions*

---

**Description**

Return CNV segmentation result from given all CNV solutions

**Usage**

```
get_segment_bysolution(solution, sample_segmentation, SL_distance_df)
```

**Arguments**

solution	solution dataframe
sample_segmentation	Segmentation dataframe
SL_distance_df	Distance matrix

**Value**

list of segmentation per solution

---

get\_solution\_table      *Get summarised table of cnv solutions*

---

**Description**

Get summarised table of cnv solutions

**Usage**

```
get_solution_table(cnv_solutions)
```

**Arguments**

cnv\_solutions    cnvcalling result from function call\_cnv.R

**Value**

Dataframe of solution table

**Examples**

```
#'  
### Loading example SampleBam file  
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")  
sample_bambin <- readRDS(example_file)  
### Example PoN  
PoN_rdsfile <- system.file("extdata", "ex.PoN.rds", package = "cfdnakit")  
pon_profiles <- readRDS(PoN_rdsfile)  
sample_profile <- get_fragment_profile(sample_bambin, sample_id = "Patient1")  
  
sample_zscore <- get_zscore_profile(sample_profile, pon_profiles)  
  
sample_zscore_segment <- segmentByPSCB(sample_zscore)  
  
sample_cnv <- call_cnv(sample_zscore_segment, sample_zscore, tfs=c(0.1,0.3), ploidy=c(1.5,2), MaxCN=3)  
get_solution_table(sample_cnv)
```

---

get\_zscore\_profile      *Transform SLRatio with PoN Fragment profile*

---

**Description**

Transform SLRatio with PoN Fragment profile

**Usage**

```
get_zscore_profile(fragment_profile, pon_profile)
```

**Arguments**

fragment\_profile      Sample Profile  
pon\_profile      PoN Profiles

**Value**

Dataframe of robust transformed SLratio

**Examples**

```
### Loading example SampleBam file
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bambin <- readRDS(example_file)

### Example PoN
PoN_rdsfile <- system.file("extdata", "ex.PoN.rds", package = "cfdnakit")
pon_profiles <- readRDS(PoN_rdsfile)
sample_profile <- get_fragment_profile(sample_bambin, sample_id = "Patient1")

sample_zscore <- get_zscore_profile(sample_profile, pon_profiles)

sample_zscore_segment <- segmentByPSCB(sample_zscore)
```

---

GRCh2UCSCGRanges

*Convert GRCh chromosome format to UCSC style*

---

**Description**

Convert GRCh chromosome format to UCSC style

**Usage**

```
GRCh2UCSCGRanges(which)
```

**Arguments**

which      GRanges object;

**Value**

GRanges; GRanges after chromosome format conversion

---

<code>if_exist_baifile</code>	<i>Check if bai file exist from given bam</i>
-------------------------------	---

---

**Description**

Check if bai file exist from given bam

**Usage**

```
if_exist_baifile(bamfile)
```

**Arguments**

<code>bamfile</code>	Character; Path to sample bamfile
----------------------	-----------------------------------

**Value**

Boolean if the bai file exist

---

<code>if_ucsc_chrformat</code>	<i>Check UCSC chromosomes format for input bam file</i>
--------------------------------	---

---

**Description**

Check UCSC chromosomes format for input bam file

**Usage**

```
if_ucsc_chrformat(bamfile_path)
```

**Arguments**

<code>bamfile_path</code>	Character; Path to sample bamfile
---------------------------	-----------------------------------

**Value**

Boolean; if the input bam file is UCSC format, chr prefix

---

make\_density\_table     *Make Fragment-length density table*

---

**Description**

Make Fragment-length density table

**Usage**

```
make_density_table(readbam_bin, minimum_length, maximum_length)
```

**Arguments**

readbam\_bin     List; A list containing SampleBam object/objects from the read\_bamfile function

minimum\_length   numeric;

maximum\_length   numeric

**Value**

data.frame

---

overlap\_bin\_with\_segment  
*Overlap and merge bin data frame with segmentation dataframe*

---

**Description**

Overlap and merge bin data frame with segmentation dataframe

**Usage**

```
overlap_bin_with_segment(per_bin_profile, sample_segmentation)
```

**Arguments**

per\_bin\_profile  
    bin dataframe

sample\_segmentation  
    segmentation dataframe

**Value**

dataframe of overlapping bin and segmentation

---

plot_cnv_solution	<i>Plot Fragment-length profile with CNV calling result</i>
-------------------	---

---

### Description

Plot Fragment-length profile with CNV calling result

### Usage

```
plot_cnv_solution(
  cnvcall,
  selected_solution = 1,
  genome = "hg19",
  ylim = c(-30, 30)
)
```

### Arguments

cnvcall	solution results from call_cnv function
selected_solution	solution rank to plot
genome	Character; version of reference genome (default hg19)
ylim	Vector of 2 Int; ylim of plot (default c(-20,20))

### Value

ggplot object plot Genomics CNV profile of selected solution

### Examples

```
### Loading example SampleBam file
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bambin <- readRDS(example_file)
### Example PoN
PoN_rdsfile <- system.file("extdata", "ex.PoN.rds", package = "cfdnakit")
pon_profiles <- readRDS(PoN_rdsfile)
sample_profile <- get_fragment_profile(sample_bambin, sample_id = "Patient1")

sample_zscore <- get_zscore_profile(sample_profile, pon_profiles)
sample_zscore_segment <- segmentByPSCB(sample_zscore)

sample_cnv <- call_cnv(sample_zscore_segment, sample_zscore, tfs=c(0.1,0.3), ploidy=c(1.5,2), MaxCN=3)
plot_cnv_solution(sample_cnv, selected_solution = 1)
```

---

plot\_distance\_matrix *Plot Distance Matrix from CNVCalling*

---

**Description**

Plot Distance Matrix from CNVCalling

**Usage**

```
plot_distance_matrix(cnvcall)
```

**Arguments**

cnvcall           cnvcalling result from function call\_cnv.R

**Value**

ggplot object ; distance matrix per cnvcalling solution

**Examples**

```
### Loading example SampleBam file
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bambin <- readRDS(example_file)
### Example PoN
PoN_rdsfile <- system.file("extdata", "ex.PoN.rds", package = "cfdnakit")
pon_profiles <- readRDS(PoN_rdsfile)
sample_profile <- get_fragment_profile(sample_bambin, sample_id = "Patient1")

sample_zscore <- get_zscore_profile(sample_profile, pon_profiles)
sample_zscore_segment <- segmentByPSCB(sample_zscore)

sample_cnv <- call_cnv(sample_zscore_segment, sample_zscore, tfs=c(0.1,0.3), ploidy=c(1.5,2), MaxCN=3)
plot_distance_matrix(sample_cnv)
```

---

plot\_fragment\_dist *Plot Fragment-length Distribution*

---

**Description**

Plot Fragment-length Distribution

**Usage**

```
plot_fragment_dist(readbam_list, maximum_length = 550, minimum_length = 20)
```

**Arguments**

readbam\_list List; A list containing SampleBam object/objects from the read\_bamfile function

maximum\_length Int; Maximum length of fragment. cfDNA fragment longer than this value will not be considered; Default 550

minimum\_length Int; Minimum length of fragment. cfDNA fragment shorter than this value will not be considered; Default 20

**Value**

distribution plot

**Examples**

```
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bambin <- readRDS(example_file)

### adding more samples to the plot
example_file2 <- system.file("extdata", "BH01_CHR15.SampleBam.rds", package = "cfdnakit")
control_bambin <- readRDS(example_file2)
readbam_list <- list(plasma1 = sample_bambin, Healthy.blood.plasma=control_bambin)
plot_fragment_dist(readbam_list)
```

---

plot\_sl\_ratio *Plot Short/Long-fragment Ratio*

---

**Description**

Plot Short/Long-fragment Ratio

**Usage**

```
plot_sl_ratio(fragment_profile, ylim = c(0, 0.4), genome = "hg19")
```

**Arguments**

fragment\_profile list

ylim plot y-axis limit

genome Character; version of reference genome (default hg19)

**Value**

plot



**Examples**

```

example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bambin <- readRDS(example_file)
sample_profile <- get_fragment_profile(sample_bambin, sample_id = "Patient1")
plot_sl_ratio(fragment_profile = sample_profile)

### change plot y-axis
plot_sl_ratio(fragment_profile = sample_profile, ylim=c(0.1,0.5))

### change reference genome
plot_sl_ratio(fragment_profile = sample_profile, genome="hg38")

```

---

plot\_transformed\_sl     *Plot z-transformed Short/Long-fragment Ratio*

---

**Description**

Plot z-transformed Short/Long-fragment Ratio

**Usage**

```

plot_transformed_sl(
  sample_transformed_sl,
  sample_segment_df = NULL,
  ylim = c(-30, 30),
  genome = "hg19"
)

```

**Arguments**

sample_transformed_sl	Dataframe z-transformed SLRatio from get_zscore_profile
sample_segment_df	Dataframe segmenation from segmentByPSCB
ylim	plot y-axis limit
genome	Character; version of reference genome (default hg19)

**Value**

Genome-wide plot of z-transformed SLRatio

**Examples**

```

### Loading example SampleBam file
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bambin <- readRDS(example_file)
### Example PoN
PoN_rdsfile <- system.file("extdata", "ex.PoN.rds", package = "cfdnakit")
pon_profiles <- readRDS(PoN_rdsfile)
sample_profile <- get_fragment_profile(sample_bambin, sample_id = "Patient1")

sample_zscore <- get_zscore_profile(sample_profile, pon_profiles)

```

```

sample_zscore_segment <- segmentByPSCB(sample_zscore)
plot_transformed_sl(sample_zscore, sample_zscore_segment)
## Change reference genome
plot_transformed_sl(sample_zscore, sample_zscore_segment, genome="hg38")

```

---

read_bamfile	<i>Read a bam file Read a bam file from give path. Alignment and sequencing read information will be binned into non-overlapping size</i>
--------------	---

---

### Description

Read a bam file Read a bam file from give path. Alignment and sequencing read information will be binned into non-overlapping size

### Usage

```

read_bamfile(
  bamfile_path,
  binsize = 1000,
  blacklist_files = NULL,
  genome = "hg19",
  target_bedfile = NULL,
  min_mapq = 20,
  apply_blacklist = TRUE
)

```

### Arguments

bamfile_path	Character; Path to sample bamfile
binsize	Int; Size of non-overlapping windows in KB. Only 100,500 and 1000 is available; Default 1000
blacklist_files	Character; Filepath to file containing blacklist regions
genome	Character; abbreviation of reference genome; available genome: hg19,hg38, mm10. default:hg19
target_bedfile	Character; Path to exon/target bedfile; Default NULL
min_mapq	Int; minimum read mapping quality; Default 20
apply_blacklist	Logical; To exclude read on the blacklist regions Default TRUE

### Value

SampleBam Object; A list object containing read information from the BAM file.

### Examples

```

f1 <- system.file("extdata","ex.plasma.bam",package = "cfdnakit")
### read bam file with default params (hg19, 1000K binsize)
sample.bam <-read_bamfile(f1, apply_blacklist=FALSE)

```

---

read_PoN_files	<i>Read Fragment Profile from a list of rds file</i>
----------------	--

---

**Description**

Read Fragment Profile from a list of rds file

**Usage**

```
read_PoN_files(list_rdsfiles)
```

**Arguments**

list\_rdsfiles path to file containing list of rds file

**Value**

list containing content of rds file

---

segmentByPSCB	<i>Segmentation data with PSCBS</i>
---------------	-------------------------------------

---

**Description**

Segmentation data with PSCBS

**Usage**

```
segmentByPSCB(sample_transformed_sl)
```

**Arguments**

sample\_transformed\_sl  
dataframe of z-transformed SLRatio

**Value**

Dataframe of segmentation result

**Examples**

```
### Loading example SampleBam file
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bambin <- readRDS(example_file)
### Example PoN
PoN_rdsfile <- system.file("extdata", "ex.PoN.rds", package = "cfdnakit")
pon_profiles <- readRDS(PoN_rdsfile)
sample_profile <- get_fragment_profile(sample_bambin, sample_id = "Patient1")

sample_zscore <- get_zscore_profile(sample_profile, pon_profiles)
sample_zscore_segment <- segmentByPSCB(sample_zscore)
```

---

```
test_ysize_KolmogorovSmirnov
      KolmogorovSmirnov test for insert size
```

---

**Description**

KolmogorovSmirnov test for insert size

**Usage**

```
test_ysize_KolmogorovSmirnov(control_insert_size, sample_insert_size)
```

**Arguments**

```
control_insert_size
      Vector of insert size of a control sample
sample_insert_size
      Vector of insert size of a testing sample
```

**Value**

KS.Test result

**Examples**

```
### Loading example SampleBam file
example_file <- system.file("extdata", "example_patientcfDNA_SampleBam.RDS", package = "cfdnakit")
sample_bambin <- readRDS(example_file)
control_rds <- "BH01_CHR15.SampleBam.rds"
control_RDS_file <- system.file("extdata", control_rds, package = "cfdnakit")
control_fragment_profile <- readRDS(control_RDS_file)
sample.isize <- extract_insert_size(sample_bambin)
healthy.isize <- extract_insert_size(control_fragment_profile)
test_ysize_KolmogorovSmirnov(sample.isize, healthy.isize)
```

---

```
UCSC2GRChSampleBam      Convert UCSC chromosome format to GRCh style from a list of alignment information
```

---

**Description**

Convert UCSC chromosome format to GRCh style from a list of alignment information

**Usage**

```
UCSC2GRChSampleBam(sample.bam)
```

**Arguments**

```
sample.bam      list of alignment information from function read_bamfile
```

**Value**

List; list of alignment information after conversion

---

util.bias_correct	<i>Correct GC Bias readcount</i>
-------------------	----------------------------------

---

**Description**

Correct GC Bias readcount

**Usage**

```
util.bias_correct(readcount, bias)
```

**Arguments**

readcount	numeric
bias	numeric

**Value**

numeric

---

zscore_transform	<i>zscore_transform transforms SLRatio profile into z-score</i>
------------------	---

---

**Description**

zscore\_transform transforms SLRatio profile into z-score

**Usage**

```
zscore_transform(per_bin_profile)
```

**Arguments**

per_bin_profile	SampleFragment from function get_fragment_profile
-----------------	---

**Value**

dataframe of z-score per bin

---

`%>%`*Pipe operator*

---

**Description**

See `magrittr::%>%` for details.

**Arguments**

<code>lhs</code>	A value or the magrittr placeholder.
<code>rhs</code>	A function call using the magrittr semantics.

**Value**

The result of calling `rhs(lhs)`.

# Index

- \* **internal**
  - %>%, [22](#)
- \* **package cf-DNA**
  - cfdnakit-package, [2](#)
  - %>%, [22](#), [22](#)
- calculate\_CES\_score, [4](#)
- call\_cnv, [5](#)
- cfdnakit (cfdnakit-package), [2](#)
- cfdnakit-package, [2](#)
- create\_blacklist\_gr, [6](#)
- create\_PoN, [6](#)
- extract\_insert\_size, [7](#)
- filter\_read\_on\_blacklist, [7](#)
- fragment\_dist, [8](#)
- get\_fragment\_profile, [8](#)
- get\_segment\_byresolution, [9](#)
- get\_solution\_table, [10](#)
- get\_zscore\_profile, [10](#)
- GRCh2UCSCGRanges, [11](#)
- if\_exist\_baifile, [12](#)
- if\_ucsc\_chrformat, [12](#)
- make\_density\_table, [13](#)
- overlap\_bin\_with\_segment, [13](#)
- plot\_cnv\_solution, [14](#)
- plot\_distance\_matrix, [15](#)
- plot\_fragment\_dist, [15](#)
- plot\_sl\_ratio, [16](#)
- plot\_transformed\_sl, [17](#)
- read\_bamfile, [18](#)
- read\_PoN\_files, [19](#)
- segmentByPSCB, [19](#)
- test\_ysize\_KolmogorovSmirnov, [20](#)
- UCSC2GRChSampleBam, [20](#)
- util.bias\_correct, [21](#)
- zscore\_transform, [21](#)