

# Package ‘crisprBowtie’

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**Version** 1.10.0

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**Title** Bowtie-based alignment of CRISPR gRNA spacer sequences

**Depends** methods

**Imports** BiocGenerics, Biostrings, BSgenome, crisprBase (>= 0.99.15),  
GenomeInfoDb, GenomicRanges, IRanges, Rbowtie, readr, stats,  
stringr, utils

**Suggests** BiocStyle, BSgenome.Hsapiens.UCSC.hg38, knitr, rmarkdown,  
testthat

**biocViews** CRISPR, FunctionalGenomics, Alignment

**Description** Provides a user-friendly interface to map on-targets and off-targets  
of CRISPR gRNA spacer sequences using bowtie. The alignment is fast,  
and can be performed using either commonly-used or custom CRISPR nucleases.  
The alignment can work with any reference or custom genomes.  
Both DNA- and RNA-targeting nucleases are supported.

**License** MIT + file LICENSE

**Encoding** UTF-8

**RoxygenNote** 7.2.1

**VignetteBuilder** knitr

**BugReports** <https://github.com/crisprVerse/crisprBowtie/issues>

**URL** <https://github.com/crisprVerse/crisprBowtie>

**git\_url** <https://git.bioconductor.org/packages/crisprBowtie>

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## Contents

runBowtie . . . . .	2
runCrisprBowtie . . . . .	3

**Index****6**


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runBowtie	<i>Perform short sequence alignment with bowtie</i>
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**Description**

Perform short sequence alignment with bowtie.

**Usage**

```
runBowtie(
  sequences,
  bowtie_index,
  bsgenome = NULL,
  n_mismatches = 0,
  all_alignments = TRUE,
  n_max_alignments = 1000,
  verbose = TRUE
)
```

**Arguments**

sequences	Character vector of DNA sequences.
bowtie_index	String specifying path to a bowtie index.
bsgenome	BSgenome object.
n_mismatches	Integer between 0 and 3 specifying maximum number of mismatches allowed between query sequences and target DNA. 0 by default.
all_alignments	Should all possible alignments be returned? TRUE by default.
n_max_alignments	Maximum number of alignments to return if all_alignments is FALSE. 1000 by default.
verbose	Should messages be printed to the console? TRUE by default.

**Details**

```
fasta <- system.file(package="crisprBowtie", "example/chr1.fa") outdir <- tempdir() Rbowtie::bowtie_build(fasta,outdir=
force=TRUE, prefix="tempIndex")
```

runBowtie can be used to map short DNA sequences to a reference genome. To search for sequences while imposing constraints on PAM sequences (such as gRNA spacer sequences), see runCrisprBowtie instead.

**Value**

A data.frame of the alignments with the following columns:

- query — string specifying query DNA sequence
- target — string specifying target DNA sequence
- chr - string specifying chromosome name
- pos - string specifying genomic coordinate of the start of the target DNA sequence
- strand - string specifying strand ("+" or "-")
- n\_mismatches - integer specifying number of mismatches between query and target sequences

**Author(s)**

Jean-Philippe Fortin

**See Also**[runCrisprBowtie](#) to map gRNA spacer sequences.**Examples**

```
fasta <- system.file(package="crisprBowtie", "example/chr1.fa")
outdir <- tempdir()
Rbowtie::bowtie_build(fasta,outdir=outdir, force=TRUE, prefix="tempIndex")
index <- file.path(outdir, "tempIndex")
seqs <- c("GGAAGT",
          "GTGGAC",
          "GTGTGC")
results <- runBowtie(seqs, bowtie_index=index, n_mismatches=2)
```

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runCrisprBowtie	<i>Perform CRISPR gRNA spacer alignment with bowtie</i>
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**Description**

Perform CRISPR gRNA spacer alignment with bowtie.

**Usage**

```
runCrisprBowtie(
  spacers,
  mode = c("protospacer", "spacer"),
  bowtie_index = NULL,
  bsgenome = NULL,
  crisprNuclease = NULL,
  canonical = TRUE,
  ignore_pam = FALSE,
  n_mismatches = 0,
  all_alignments = TRUE,
  n_max_alignments = 1000,
  force_spacer_length = FALSE,
  rna_strict_directionality = TRUE,
  verbose = TRUE
)
```

**Arguments**

spacers	Character vector specifying gRNA spacer sequences. Sequences must all be of equal length.
mode	String specifying which alignment mode should be used: protospacer or spacer. For RNA-targeting nucleases such as CasRx, only the protospacer mode can be used.

bowtie_index	String specifying path to a bowtie index.
bsgenome	A <a href="#">BSgenome</a> object. Must be provided if mode is "spacer". Ignore
crisprNuclease	A <a href="#">CrisprNuclease</a> object.
canonical	Should only canonical PAM sequences be considered? TRUE by default.
ignore_pam	Should PAM sequences be ignore? If TRUE, all alignments are returned regardless of PAM tolerance. FALSE by default.
n_mismatches	Integer between 0 and 3 specifying maximum number of mismatches allowed between spacer sequences and target DNA. 0 by default.
all_alignments	Should all possible alignments be returned? TRUE by default.
n_max_alignments	Maximum number of alignments to return if all_alignments is FALSE. 1000 by default.
force_spacer_length	Should the spacer length be overwritten in the <code>crisprNuclease</code> object? FALSE by default.
rna_strict_directionality	Should only protospacers found in the original direction of the RNA be considered for RNA-targeting nucleases? TRUE by default.
verbose	Should messages be printed to the console? TRUE by default.

## Details

When mode is "spacer", spacer sequences are aligned to the reference index without appending PAM sequences first. This requires the specification of a [BSgenome](#) object through the argument `bsgenome` to validate that the aligned spacer sequences are adjacent to valid PAM sequences.

When mode is "protospacer", sequences are aligned with all valid PAM sequences appended (spacer + PAM). The set of valid PAM sequences depend on the inputs `canonical` and `ignore_pam`. This is faster than the "spacer" mode if the number of possible PAM sequences is small (e.g. SpCas9).

For RNA-targeting nucleases, such as RfxCas13d (CasRx), the bowtie index should be built on a transcriptome. For such applications, only the "protospacer" mode can be used as there is no corresponding `bsgenome` package. The protospacer sequences searched in the reference index will be the reverse complement of the input spacer sequences.

## Value

A data.frame of the spacer alignments with the following columns:

- `spacer` — string specifying gRNA spacer sequence
- `protospacer` — string specifying target protospacer sequence
- `pam` — string specifying target PAM sequence
- `chr` - string specifying chromosome name
- `pam_site` - string specifying genomic coordinate of the first nucleotide of the PAM sequence.
- `strand` - string specifying strand ("+" or "-")
- `n_mismatches` - integer specifying number of mismatches between spacer and protospacer sequences
- `canonical` - logical indicating whether or not PAM sequence is canonical.

**Author(s)**

Jean-Philippe Fortin

**See Also**

[runBowtie](#) to map general DNA sequences.

**Examples**

```
fasta <- system.file(package="crisprBowtie", "example/chr1.fa")
outdir <- tempdir()
Rbowtie::bowtie_build(fasta,outdir=outdir, force=TRUE, prefix="tempIndex")
index <- file.path(outdir, "tempIndex")
seqs <- c("GGAAATCCCCCAGTGGCGC",
          "ACACAGCTGCGGACAGGGCC")
data(SpCas9, package="crisprBase")
results <- runCrisprBowtie(seqs,
                           bowtie_index=index,
                           n_mismatches=2,
                           crisprNuclease=SpCas9)
```

# Index

BSgenome, [4](#)

CrisprNuclease, [4](#)

runBowtie, [2](#), [5](#)

runCrisprBowtie, [3](#), [3](#)