# Introduction to pre-processing – lab

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This lab introduces pre-processing and quality assessment, focusing on Affymetrix single channel data. It is based on Chapter 3 of *Bioconductor Case Studies*, by Hahne, Huber, Gentleman, and Falcon.

1. Briefly read through the chapter 3 handout.
2. Ensure that you have the CLL package installed. To do this, start an R session and type the command

   ```
   > library(CLL)
   ```

   If R responds with `Error in library(CLL) : there is no package called 'CLL'` then use the ethernet connection to install the package:

   ```
   > source("http://wilson2.fhcrc.org/installScripts/biocLite.R")
   > biocLite(CLL)
   ```

   (use `source("http://bioconductor.org/biocLite.R")` for access after the course is complete.)
3. (Optional) Copy the CEL files from the thumb drive to your disk.

**Exercise 1**
*Attach the CLL package, and use the* `data` *function to load* `CLLbatch` *into your work space. The* `CLLbatch` *object contains raw probe values; they have not been summarized or normalized in any way. What is the class of* `CLLbatch`*? Can you find the help page describing this class? What about a help page describing the* `CLLbatch` *itself? From looking at what is printed when you type* `CLLbatch` *at the command line, what basic information (e.g., number of genes, number of samples, the chip on which the expression data were collected) can you obtain about* `CLLbatch`*?*

**Exercise 2**
*As an exercise, we'll add some phenotypic data* `CLLbatch`*. Do this by using* `data(disease)`*. Take a minute to explore this object. What class is it, how many rows does it have, what are its column names?*

*Update the row names of the phenotype data.frame to reflect the SampleID, and remove the* `.CEL` *file extension from the sampleNames of the AffyBatch.*

*Create a vector* `mt` *that tells us how the row names of the phenotype data.frame align with the column names of the AffyBatch.*

*Create the 'metadata' describing the phenotype data, then an AnnotatedDataFrame combing the phenotype data and metadata, and finally add this to the* `CLLbatch` *AffyBatch. Use* `mt` *to make sure the order of the phenotype and expression data is consistent.*

*One array is missing disease status. Drop this from further analysis.*

**Exercise 3**

*There is an array quality metrics report for* `CLLbatch`*; we'll review it in class. The report suggests that array CLL1 is suspect, so remove it from further analysis.*

**Exercise 4**

*Pre-process the AffyBatch using* `rma`*. What is the class of the resulting object? How many probesets are there in this object? Can you extract the summarized expression values (using the* `exprs` *function? What's happened to the phenotypic data that we added to* `CLLbatch`*?*

*Often* `just.rma` *is used to go directly from CEL file to ExpressionSet in a fast and memory efficient way.*

**Exercise 5**

*Functions can often help to reduce the need to repeatedly perform routine tasks, and in the process make work flow less error prone. The NUSE boxplot in the chapter represents two steps: processing an AffyBatch object using* `fitPLM`*, and then plotting the results. Capture these steps in a function, and add it to your package.*

*Some steps might include:*

1. *Decide on what your function will do: for an AffyBatch object, fit a probewise linear model. Then create a NUSE plot.*

2. *Decide what the inputs and outputs of your function must be. The input will be an AffyBatch object. The 'output' is really a side effect, the creation of a plot.*

3. *Create a simple function taking the appropriate inputs and outputs, but doing nothing else. Choose a function name and arguments with an eye toward making their purpose clear. Create the function in a file in the* `R` *directory of your simple package.*

4. *Fill in the function with the details.*

5. *Test the function.*