

Exercises: Using Bioconductor Annotations

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These exercises will take us through various kinds of practical examples to make sure that you are comfortable using Bioconductor annotations.

Pre-requisite: The *Annotations* package successfully installed and attached.

```
> install.packages("Annotations_1.0.4.tar.gz", repos=NULL,  
+                  type="source")
```

Exercise 1

Load the following toy example of a *topTable* from the *Annotations* package:

```
> library(Annotations)  
> load(system.file("data", "tt.Rda", package="Annotations"))  
> tt
```

	ID	logFC	t	P.Value	adj.P.Val	B
1	100127974	1.6665129	6.236165	0.0001280333	0.01280333	1.4677893
2	10013	1.6159731	4.818758	0.0008459081	0.04229540	-0.4563193
30	100130000	0.9324490	3.564729	0.0056950215	0.15242524	-2.3996197
29	100130001	-0.9453525	-3.522244	0.0060970095	0.15242524	-2.4686231
86	100130002	0.7177127	2.788740	0.0203227814	0.34371907	-3.6739124
50	100130003	-0.9524966	-2.779932	0.0206231440	0.34371907	-3.6884049
98	100130004	-0.7254350	-2.530065	0.0312897608	0.43511296	-4.0974726
9	100130006	0.4959552	2.466192	0.0348090364	0.43511296	-4.2011172
63	100130009	-1.6781056	-2.343702	0.0426907899	0.47434211	-4.3983413
88	100130011	0.6942891	2.198697	0.0543065097	0.50212044	-4.6285126

Now find the gene *Symbol* and *pubmed IDs* for the top gene. Then use the *pubmed ID* that turns up to find other genes that were associated with that publication.

Exercise 2

Modify the *topTable* to include the gene symbols and chromosomes that match with the gene IDs.

Exercise 3

Get the *GO terms* for the 2nd most relevant gene from the *topTable*.

Exercise 4

Load the `transcriptDb` package for `TxDb.Hsapiens.UCSC.hg19.knownGene.db`. And then apply a filter on the chromosomes so that only chromosome 7 is exposed. Finally, extract the transcripts into a `GRangesList` object grouping by gene.

Exercise 5

The following will load a partial `topTable` such as you might get from the `DEXseq` package.

```
> load(system.file("data",
+                 "ttDEX.Rda",
+                 package="Annotations"))
```

Notice that this table has both `entrez gene IDs` and `exon IDs`, use these to find 1) the gene symbols that correspond to the various elements, 2) the ranges for the corresponding transcripts and 3) the ranges for the corresponding exons.

Exercise 6

Read in a gapped alignment using the code below:

```
> ga <- readGappedAlignments(system.file("extdata",
+                                       "chr7Cont1.bam",
+                                       package="Annotations"))
```

Now take that data, and the annotation data for the transcripts and use `countOverlaps` to determine how many reads are aligned with the "CFTR" (cystic fibrosis) gene.

Exercise 7

Begin this exercise by making a `FeatureDb` from the `oreganno` table in the `oreganno` track. Next, change the chromosome filtering on the `TranscriptDb` that we loaded earlier for `hg18` so that it uses the standard set of human chromosomes (`chr1:chr22`, plus `chrM`, `chrX` and `chrY`). Then use the `matchMatrix` object produced by `findOverlaps()` to determine which `oreganno` elements overlap with transcripts in the `TxDb`. Determine which of the genes had the most overlapping elements and then look up the gene symbol for it.