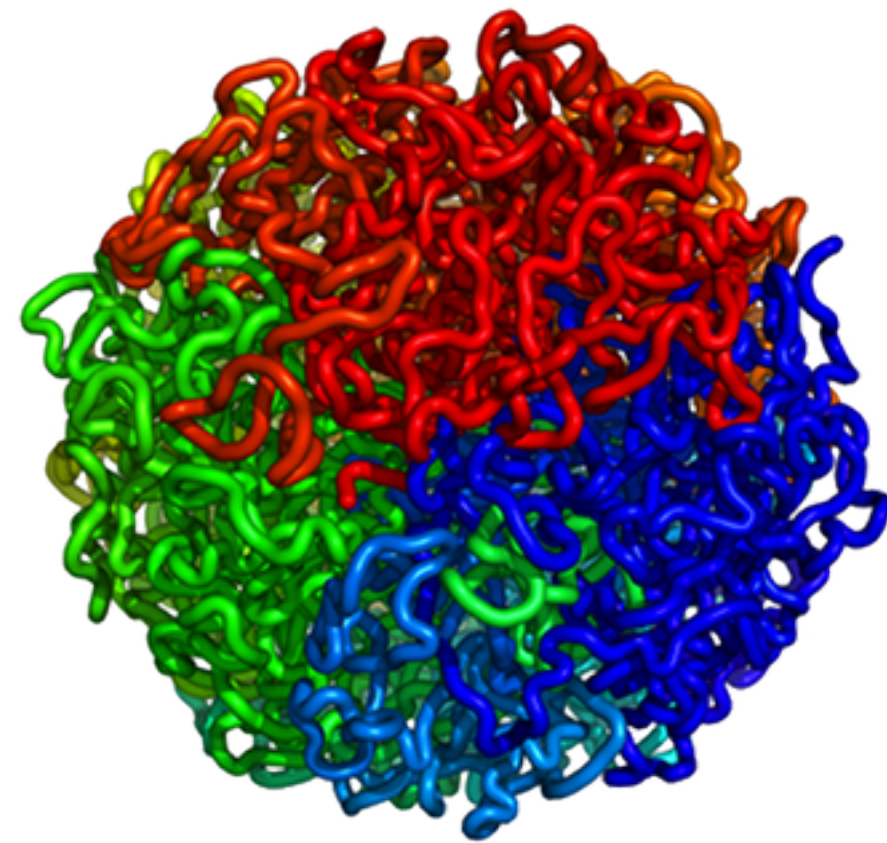


Analyzing 3-D genome organization

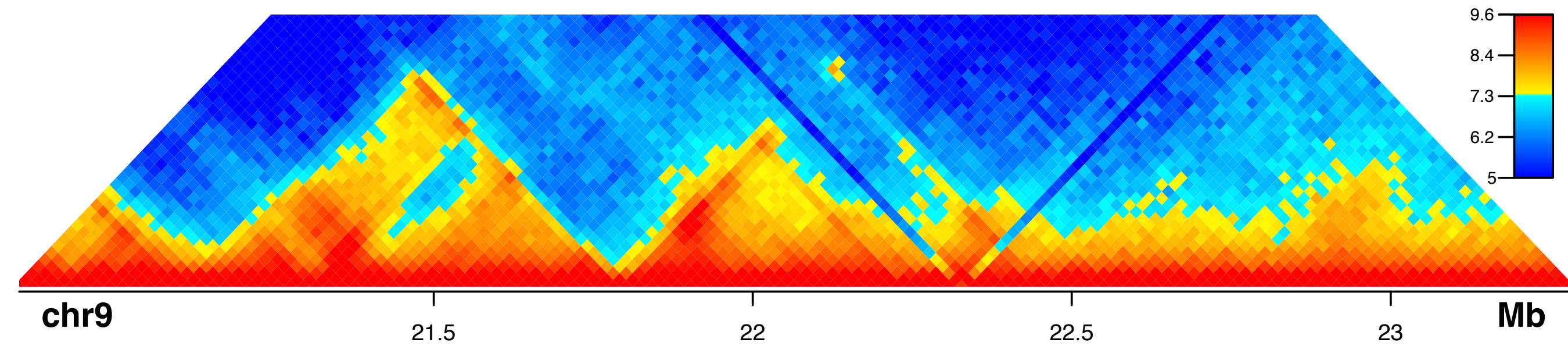
Martin Aryee

Massachusetts General Hospital
Harvard T.H. Chan School of Public Health
Broad Institute

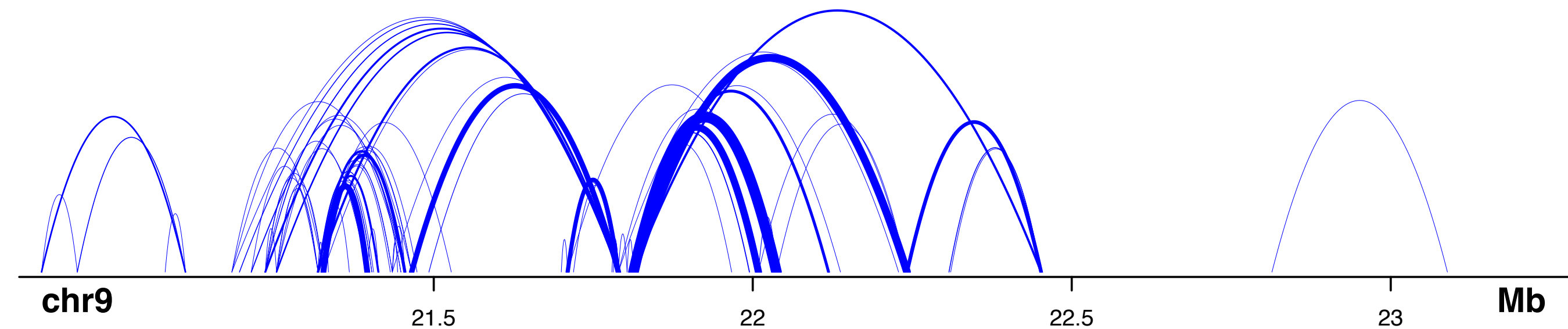


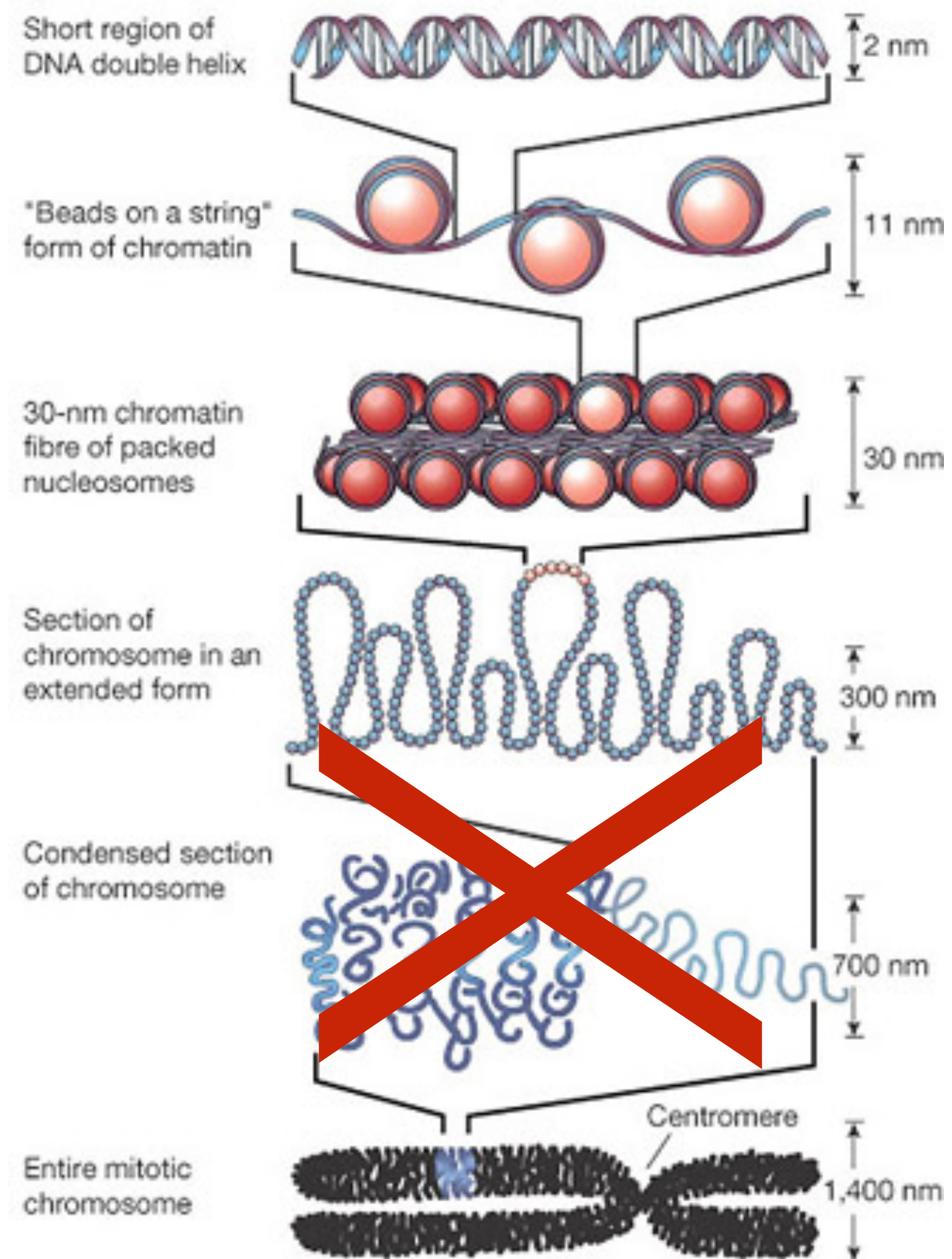
<http://www.aidenlab.org>

GM12878-HiC

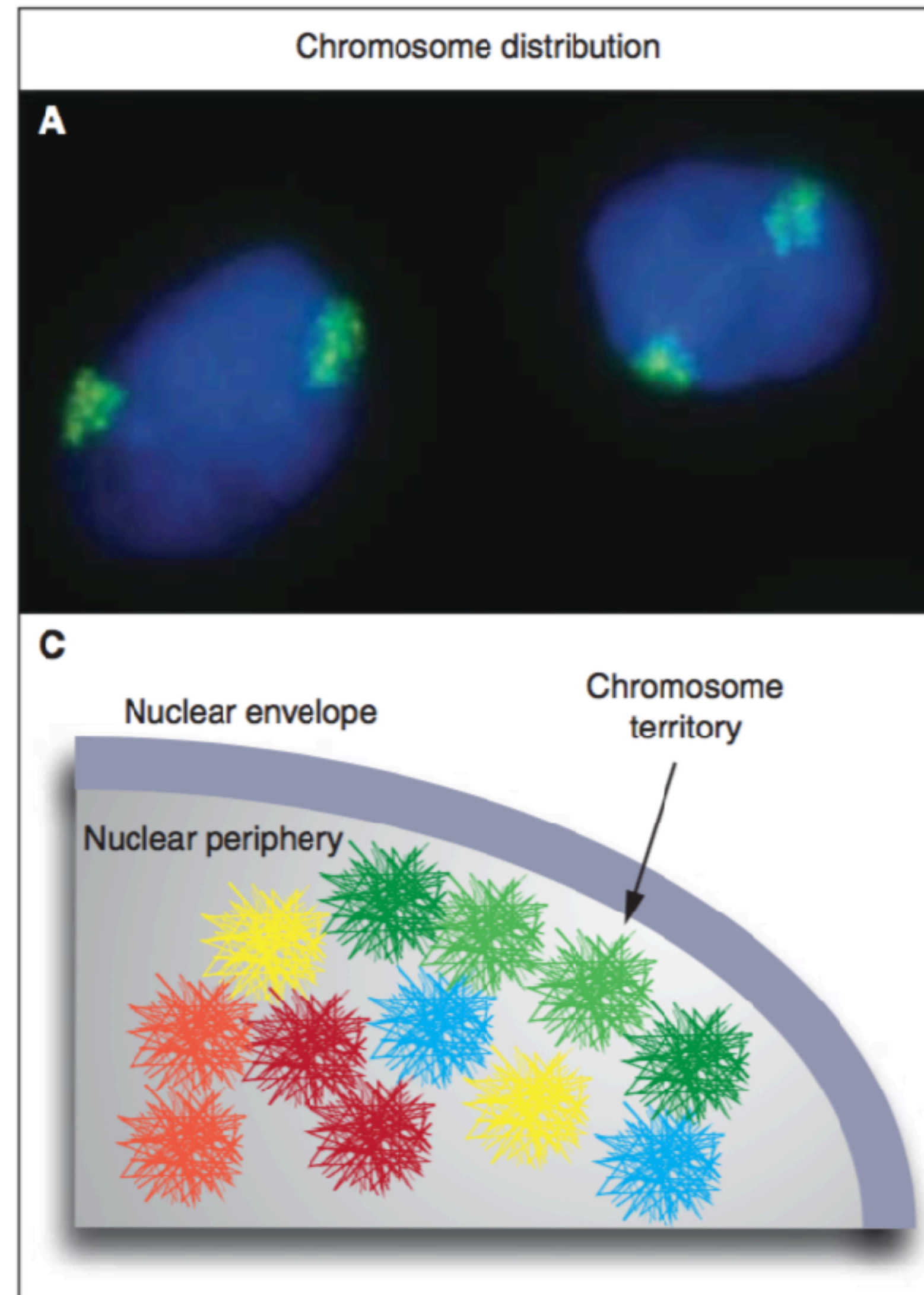


GM12878-ChIA-Pet-CTCF





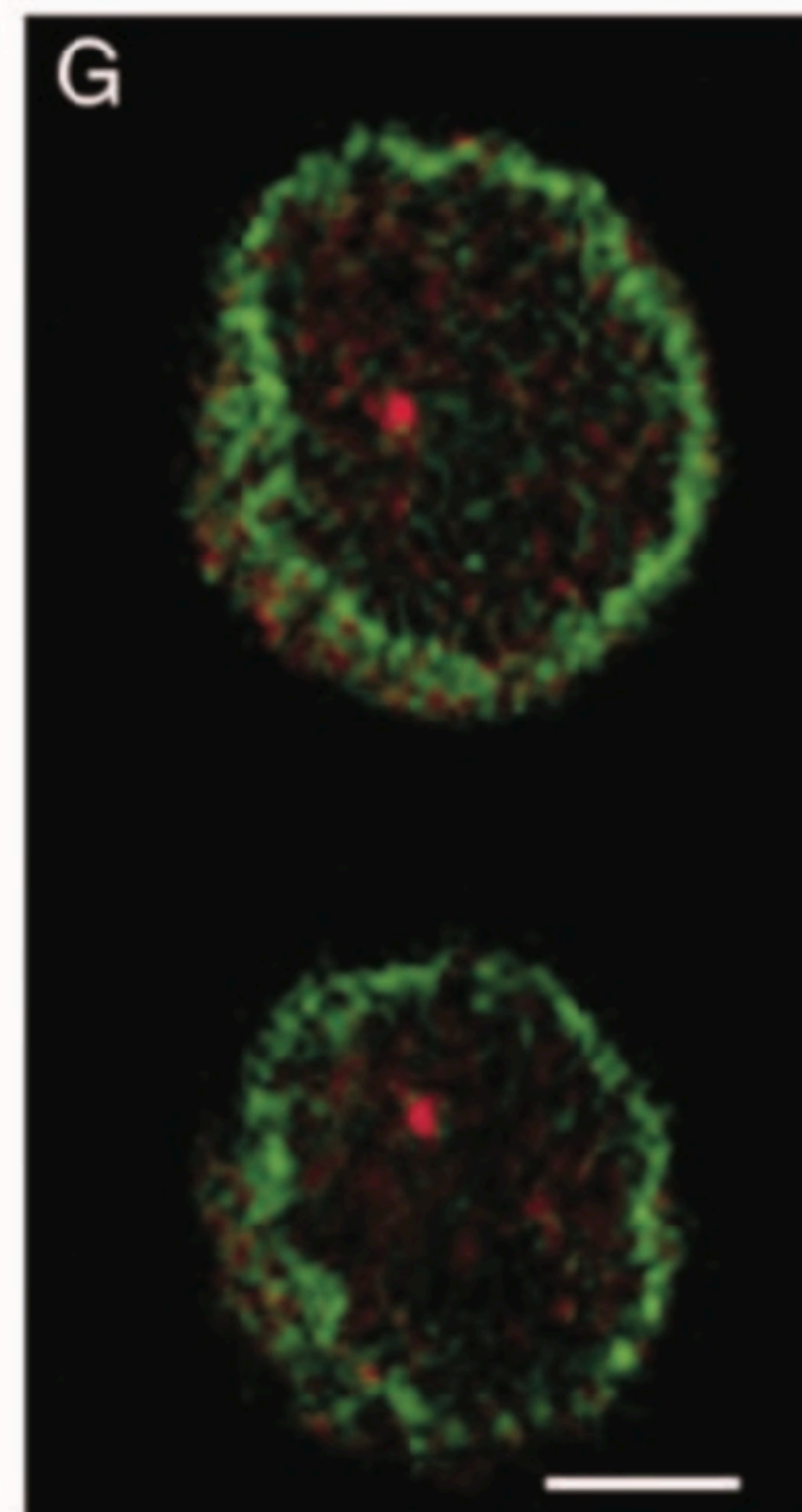
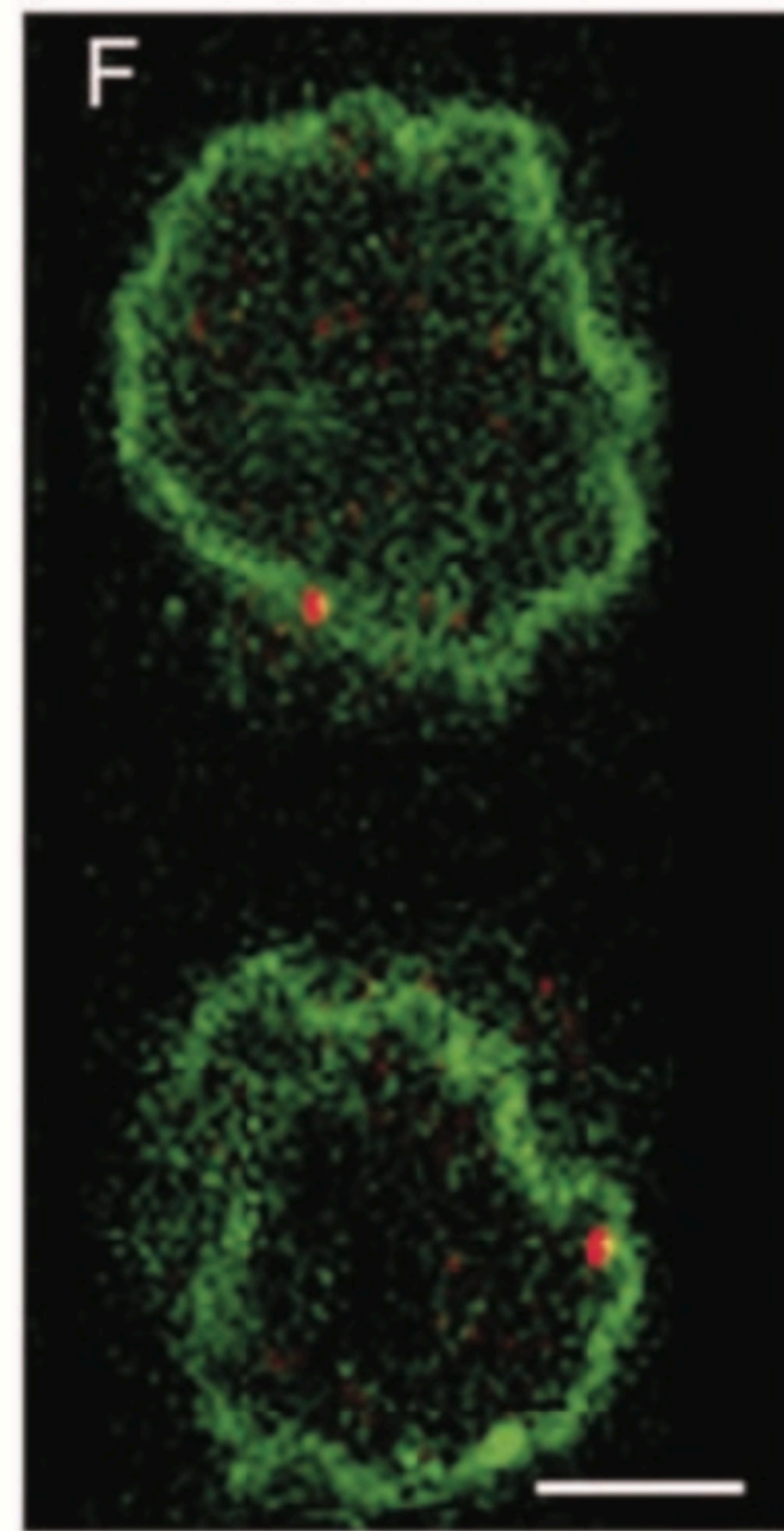
Chromosomes exhibit non-random spatial organization



Gene repositioning is associated with gene activation

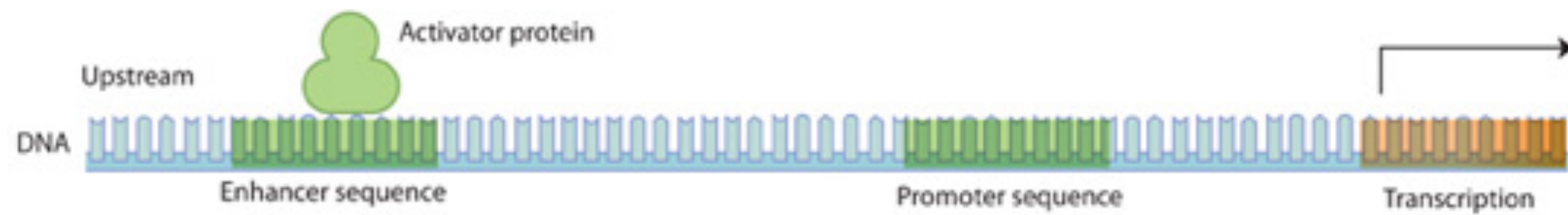
Silenced gene

Active, expressed gene

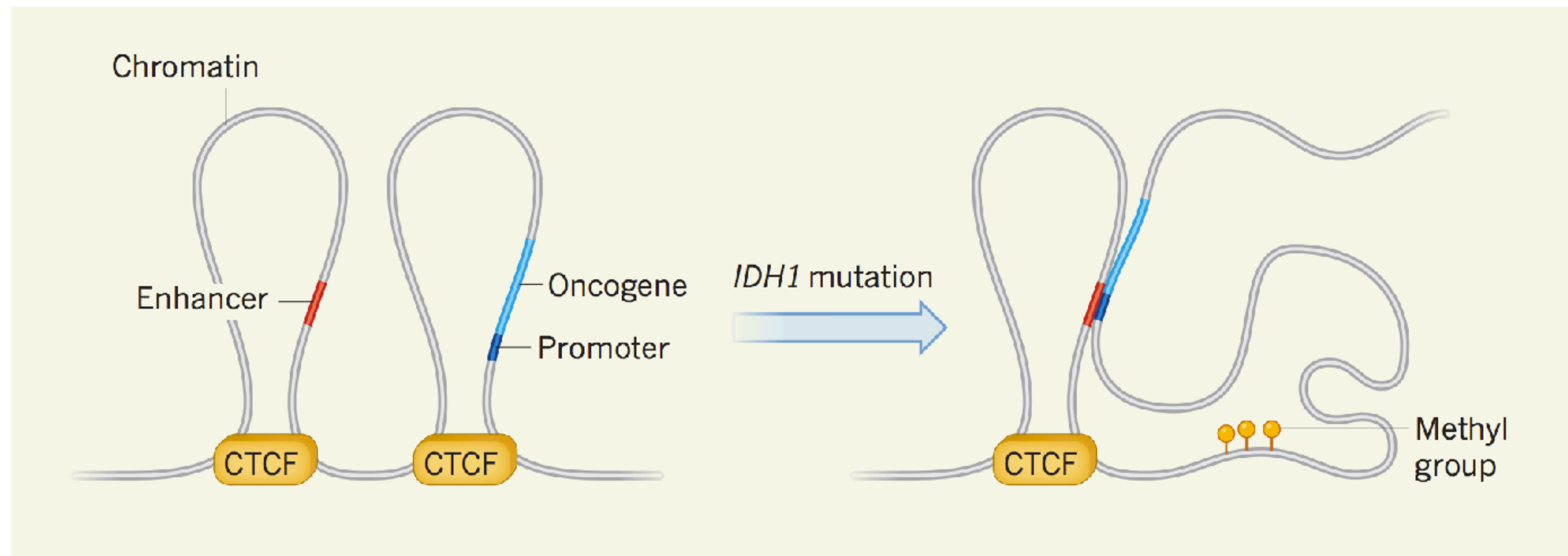


Green = anti-lamin B
Red = IgH locus

Enhancer - promoter looping



Disruption of genome topology in cancer

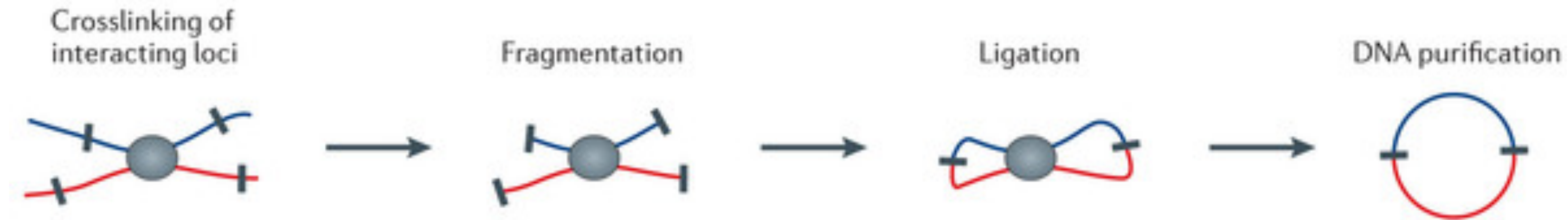


Flavahan and Drier et al., Nature (2015)
Image from Grimmer & Costello, Nature (2015)

“Structural” vs. “Functional” loops

Genome topology assays

a 3C: converting chromatin interactions into ligation products



b Ligation product detection methods

3C	4C	5C	ChIA-PET	Hi-C
One-by-one All-by-all	One-by-all	Many-by-many	Many-by-many	All-by-all
			<ul style="list-style-type: none"> • DNA shearing • Immunoprecipitation 	<ul style="list-style-type: none"> • Biotin labelling of ends • DNA shearing
PCR or sequencing	Inverse PCR sequencing	Multiplexed LMA sequencing	Sequencing	Sequencing

Genome-wide

~200M-500M Reads

~1-5 Billion Reads

QC: Hi-C

- Key metric for both Hi-C and ChIA-PET:

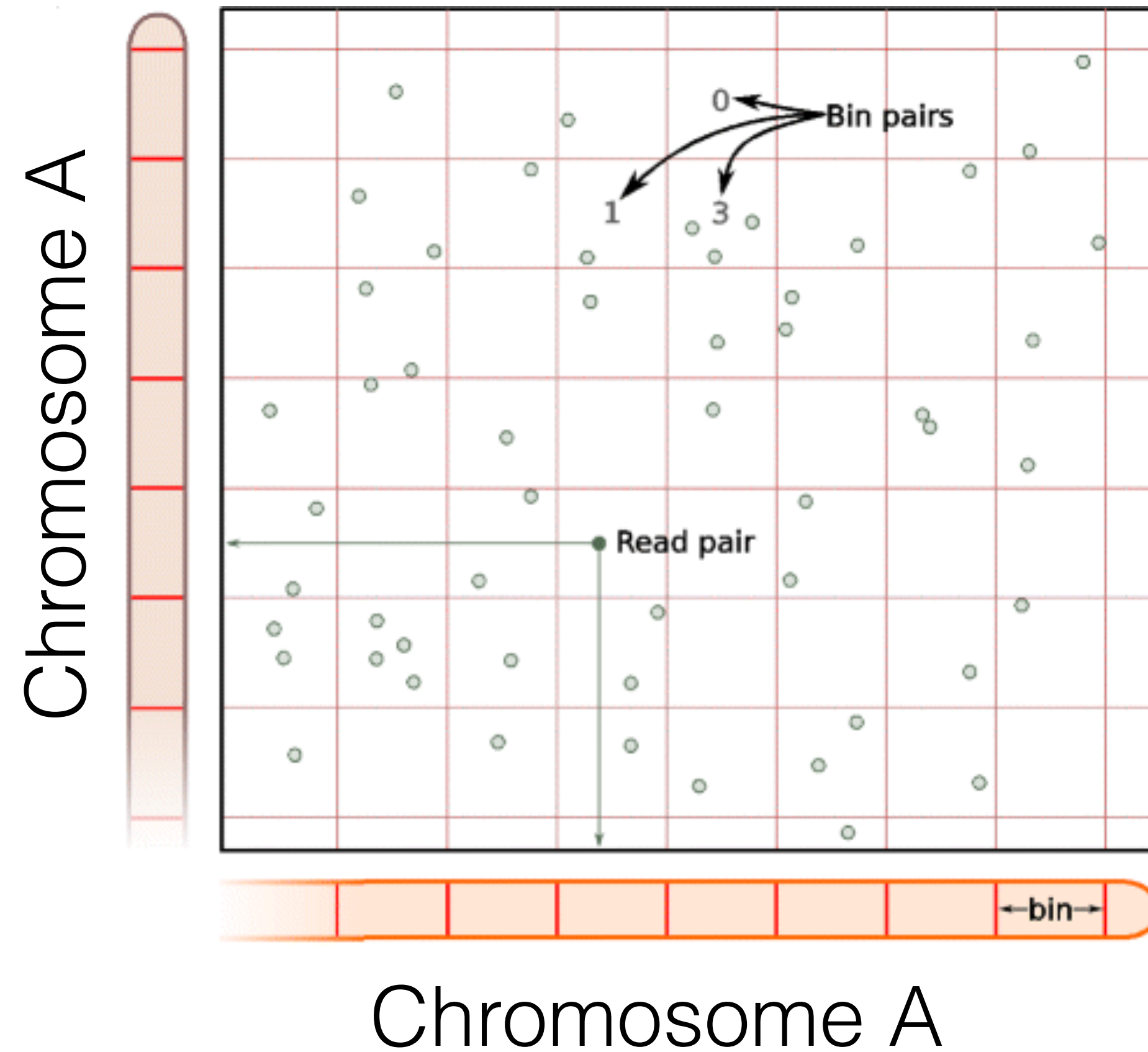
Fraction of paired-end read (“PETs”) supporting long-range interactions



https://www.bioinformatics.babraham.ac.uk/projects/hicup/scripts_description/

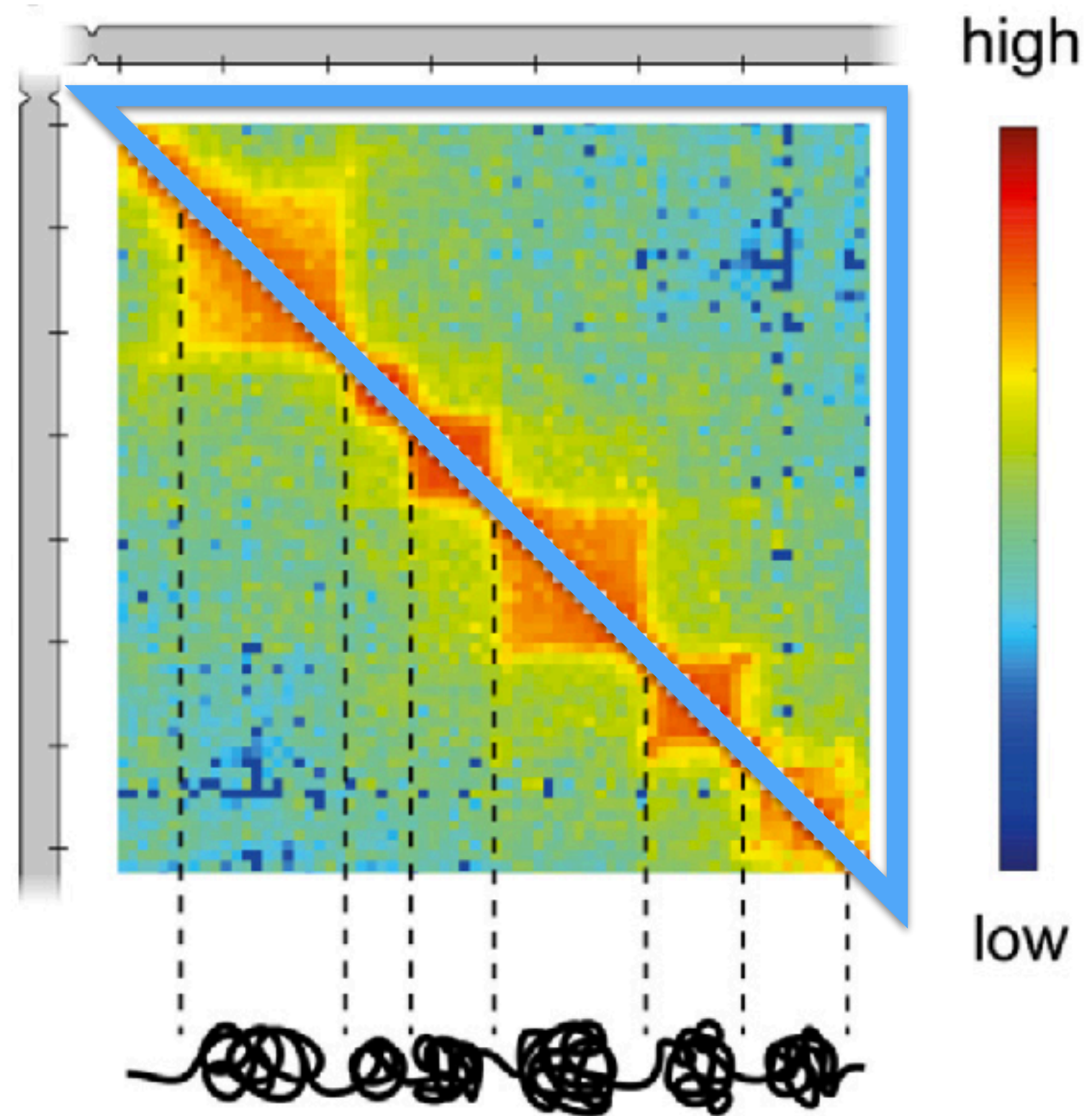
- Poor Hi-C library: <20% long-range (>20kb) pairs
- Good Hi-C library: ~40% long-range (>20kb) pairs
- A shallow sequencing run (~5M reads) is sufficient to assess library quality

HiC data: From read pairs to an interaction matrix

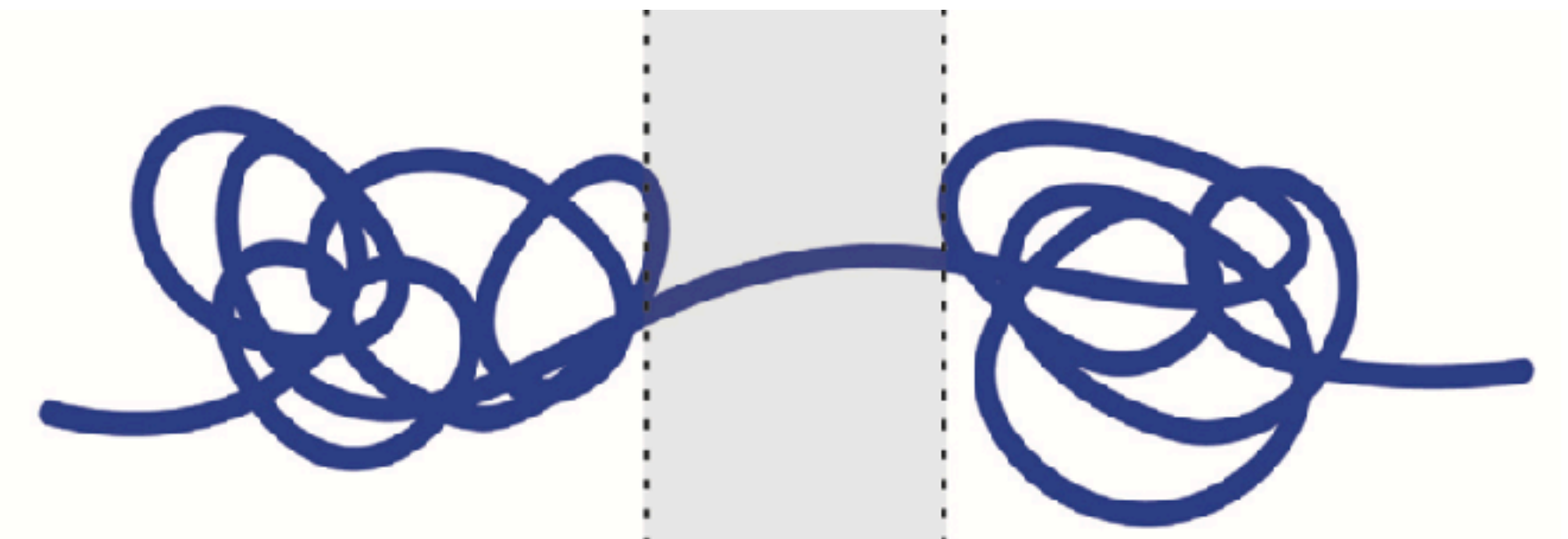


Resolution (bin size)	Number of matrix cells
1 Mbase	~10M
10 kbase	~100B

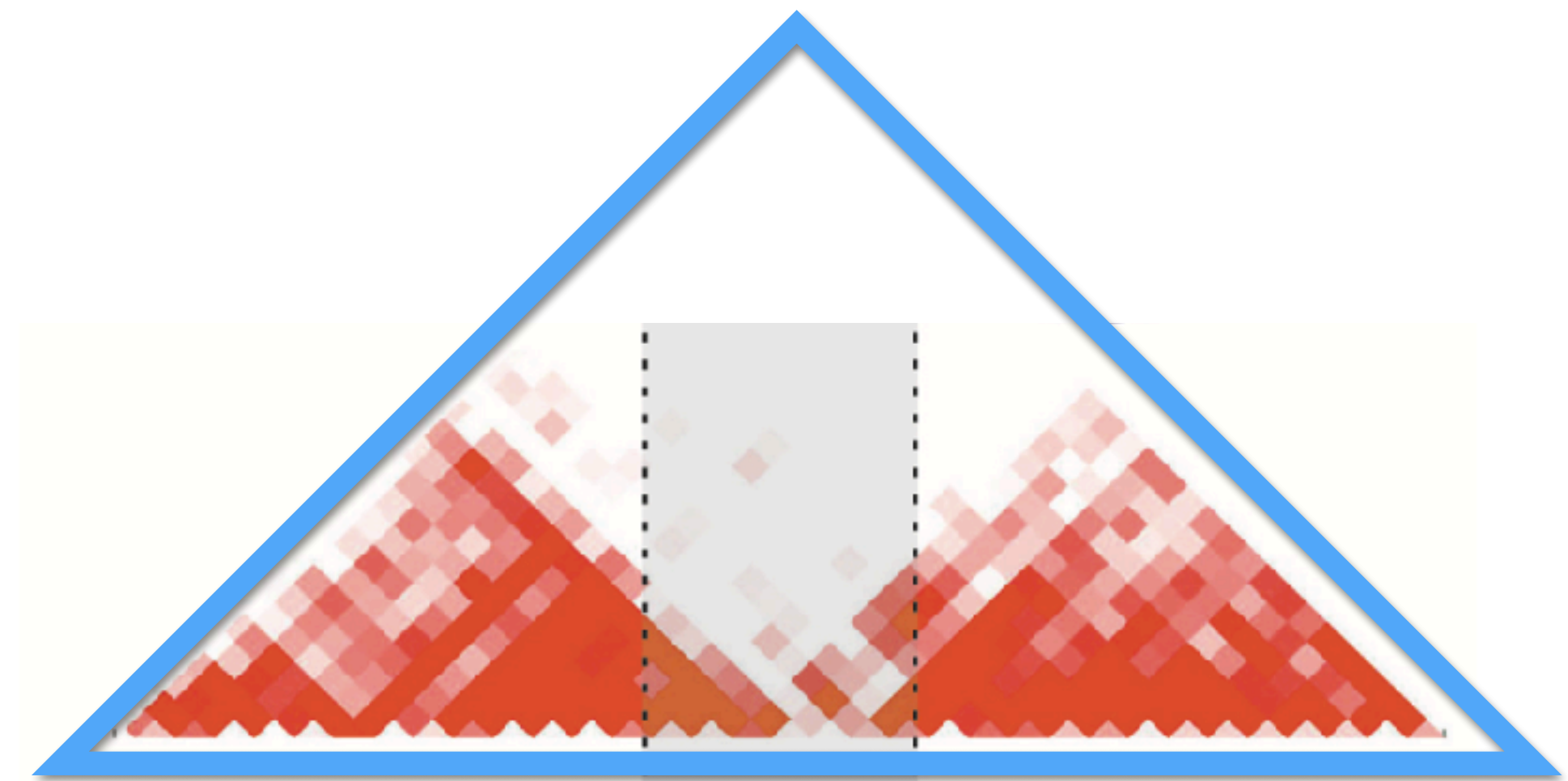
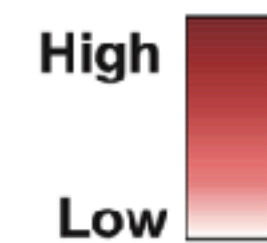
HiC data: From read pairs to an interaction matrix



Chromatin architecture



Interaction frequency (Hi-C)

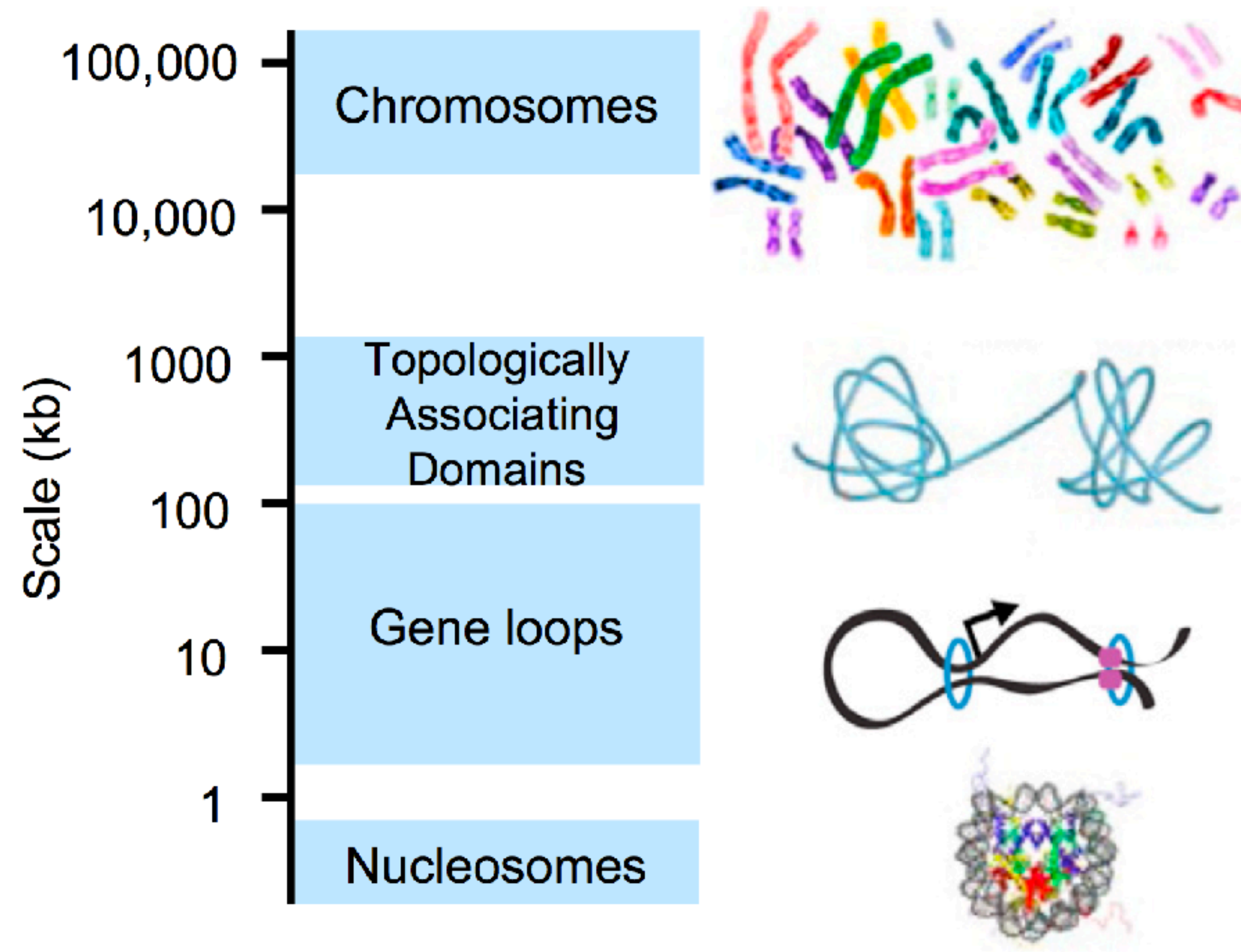


From Ulianov, S.V., Gavrilov, A.A., Razin, S.V., 2015. Nuclear Compartments, Genome Folding, and Enhancer-Promoter Communication. In: Jeon, K.W. (Ed.), International Review of Cell and Molecular Biology, pp. 183–244.
ISBN: 9780128022825
Copyright © 2015 Elsevier Inc. All rights reserved.
Academic Press

Dixon, J. R., Gorkin, D. U. & Ren, B. Chromatin Domains: The Unit of Chromosome Organization. Molecular Cell 62, 668–680 (2016).

3D genome organization at multiple scales

A



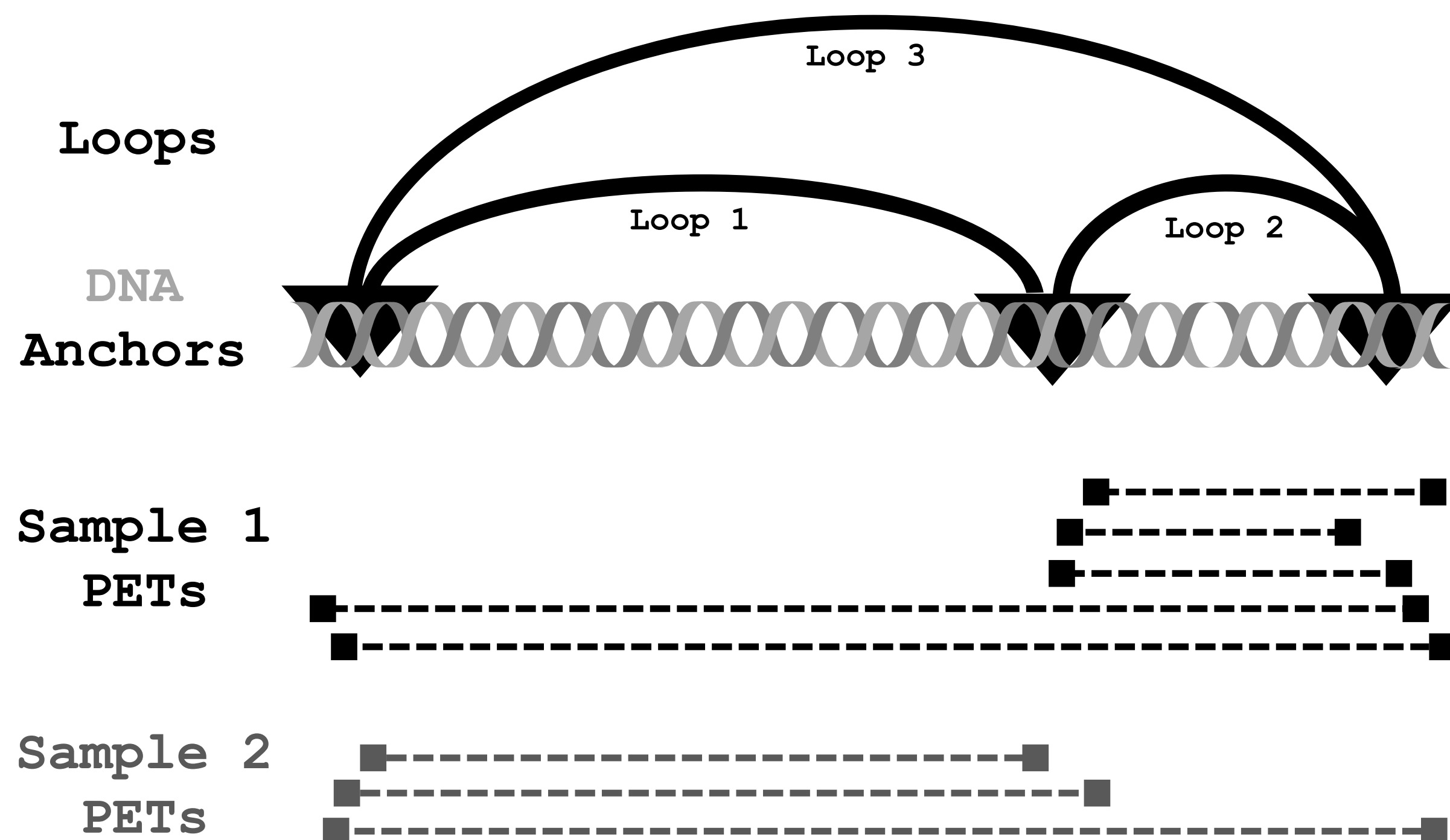
HiChIP / ChIA-PET data:

From read pairs to an interaction matrix



Processing steps:

1. Identify loop anchors (i.e. ChIP peaks)
2. Anchor pairs joined by PETs are putative loops
3. Count PETs (paired-end reads) per putative loop
4. Determine loop significance (i.e. is the number of PETs higher than the background?)

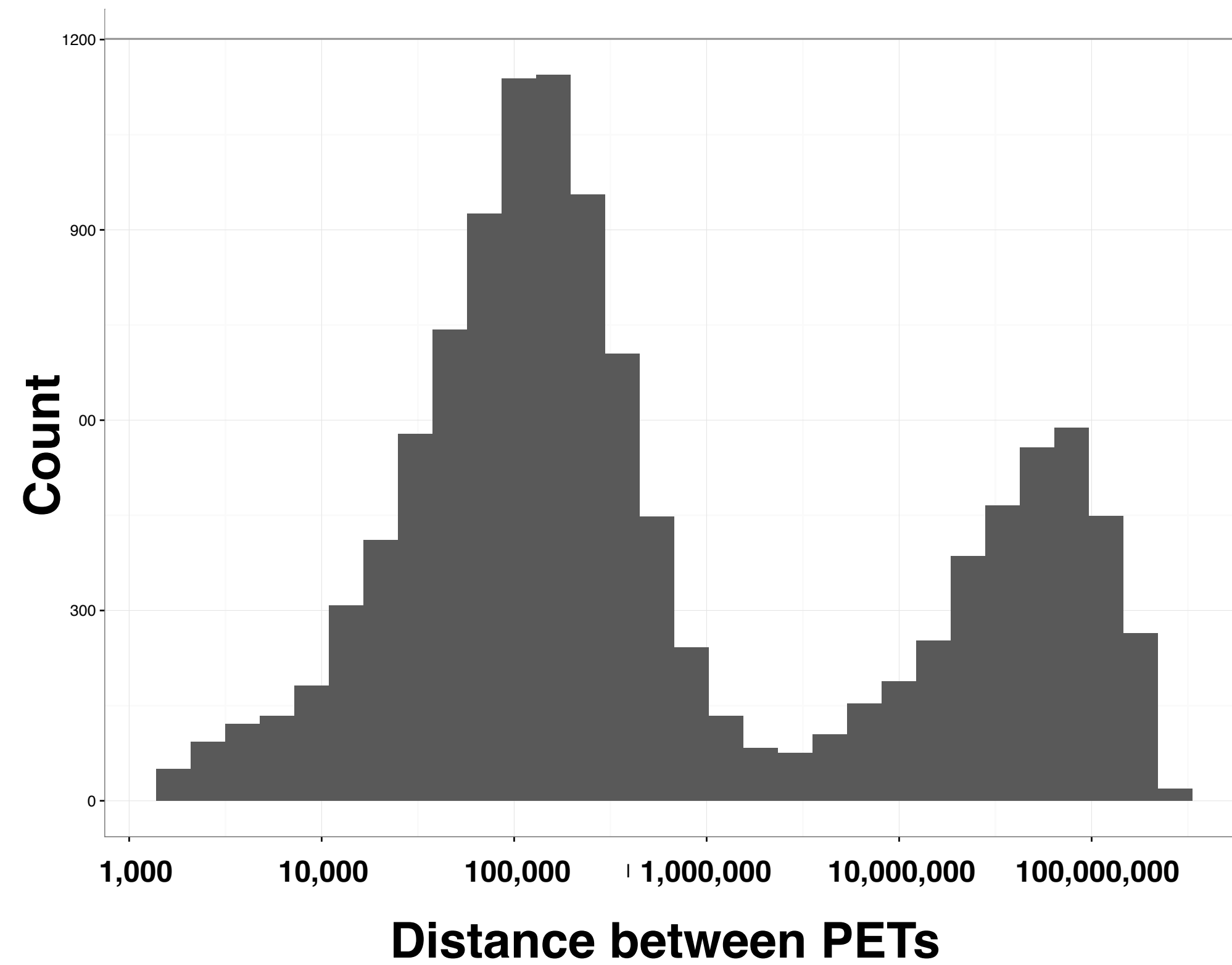


	Sample 1	Sample 2
Loop 1	0	2
Loop 2	3	0
Loop 3	2	1

PET = Paired End Tag



How long are chromatin loops?

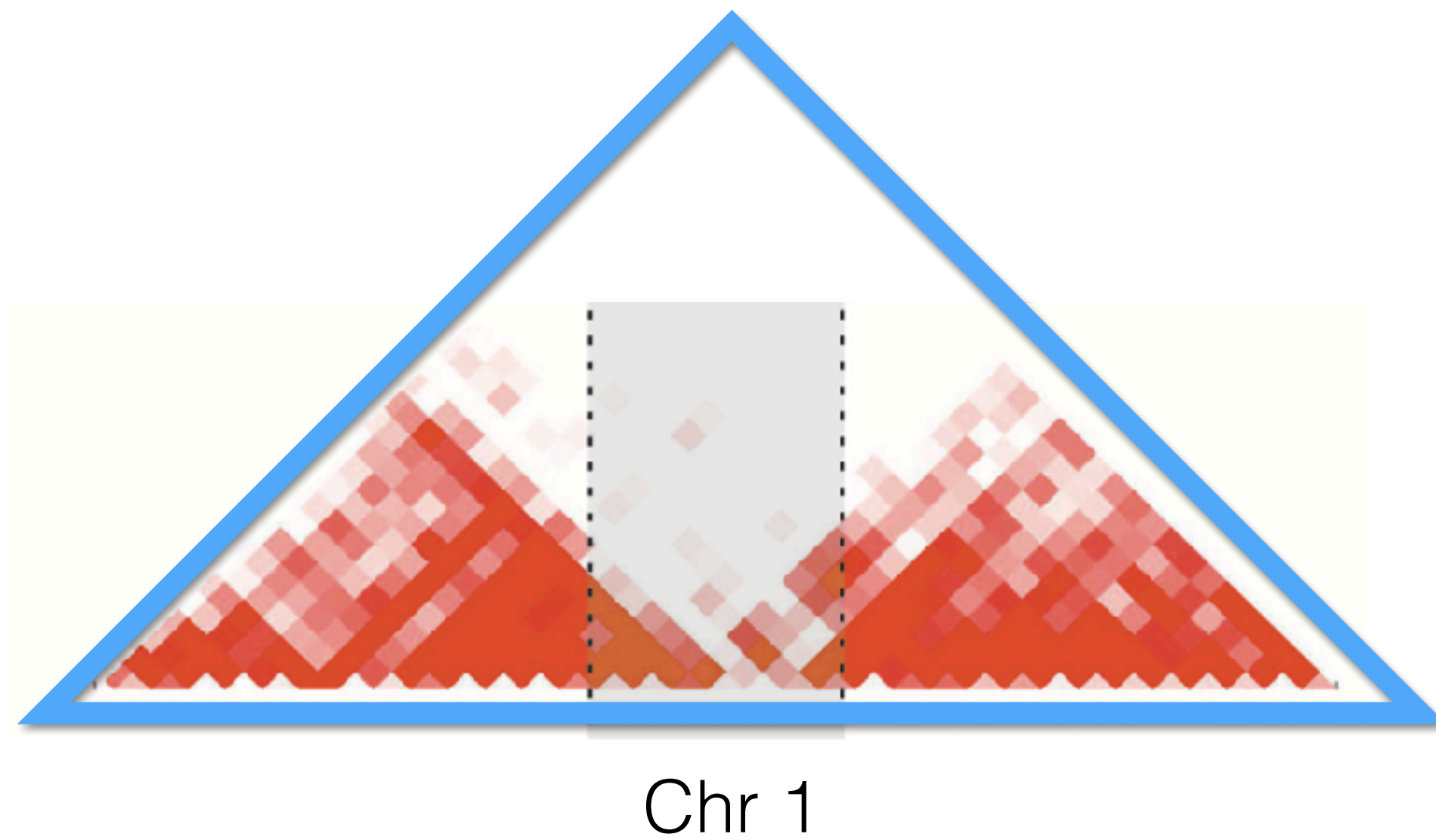


Hi-C

All pairwise interactions

Interaction
frequency
(Hi-C)

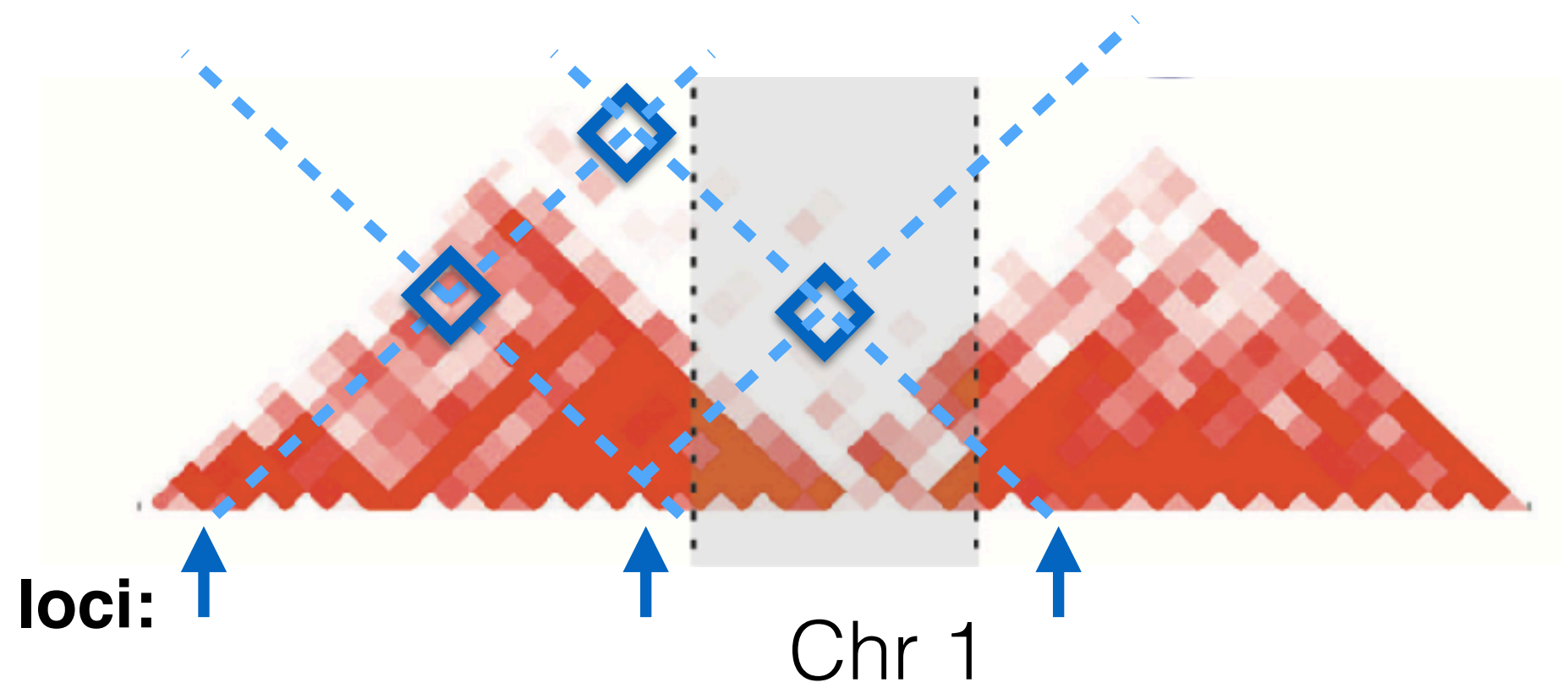
High
Low



ChIA-PET

Pairwise interactions between
loop anchor loci

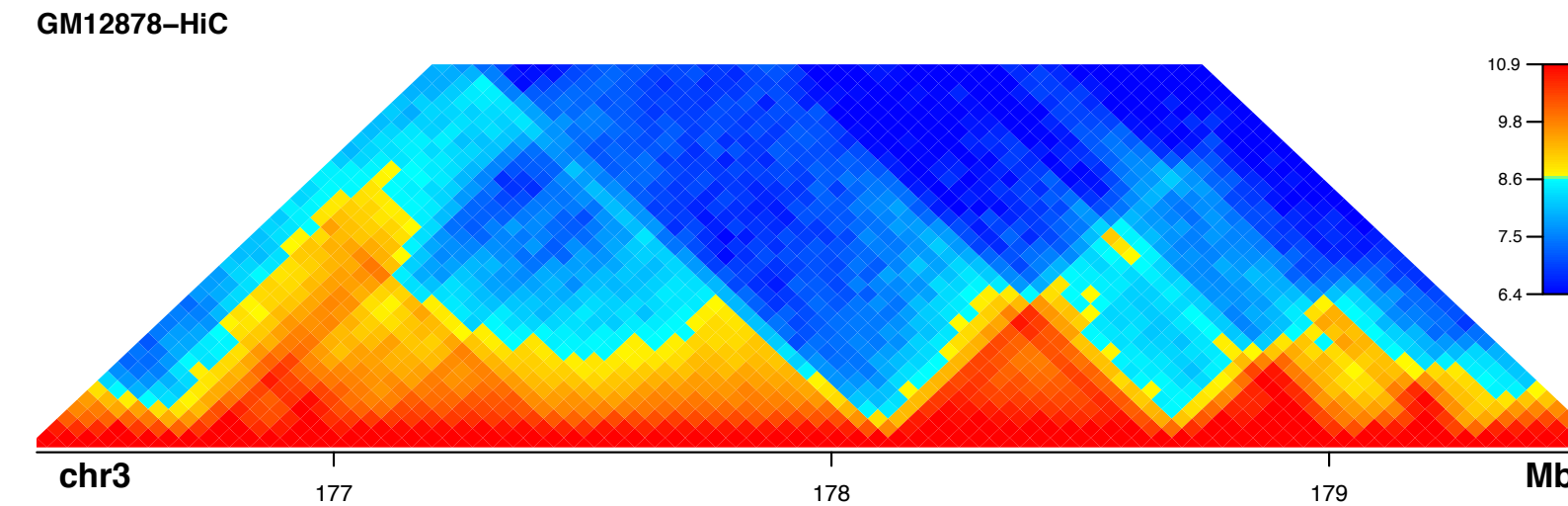
Loop anchor loci:



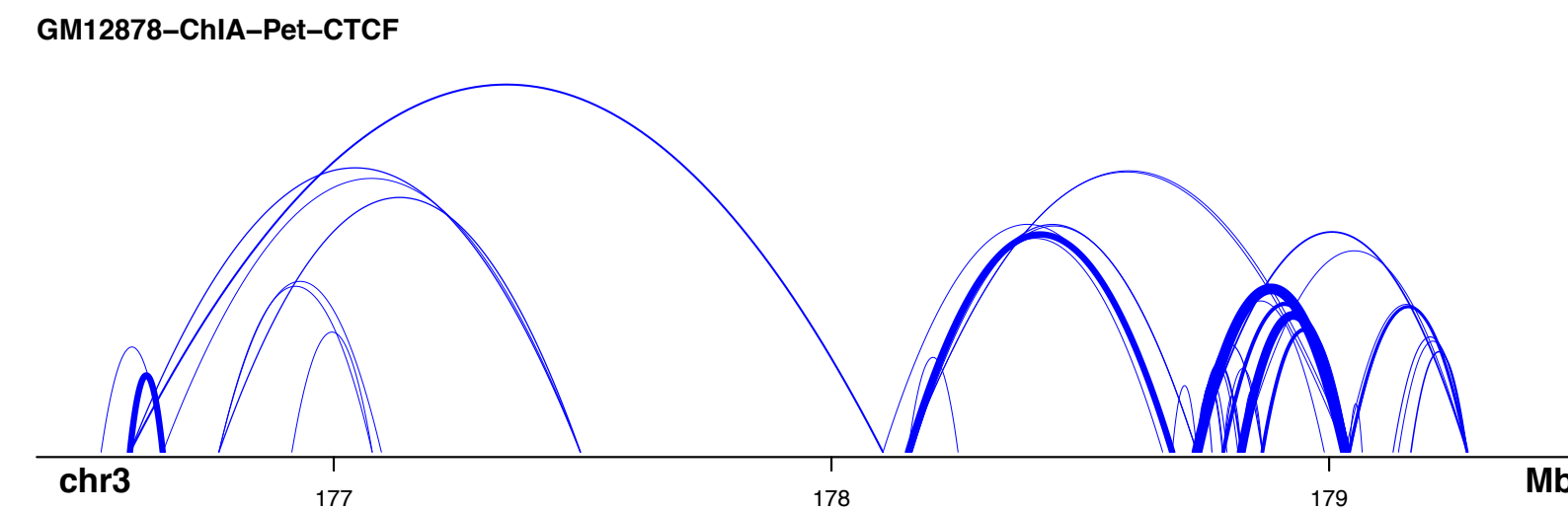
Adapted from: Dixon, J. R., Gorkin, D. U. & Ren, B. Chromatin Domains: The Unit of Chromosome Organization. *Molecular Cell* 62, 668–680 (2016).

Hi-C and ChIA-PET are complementary

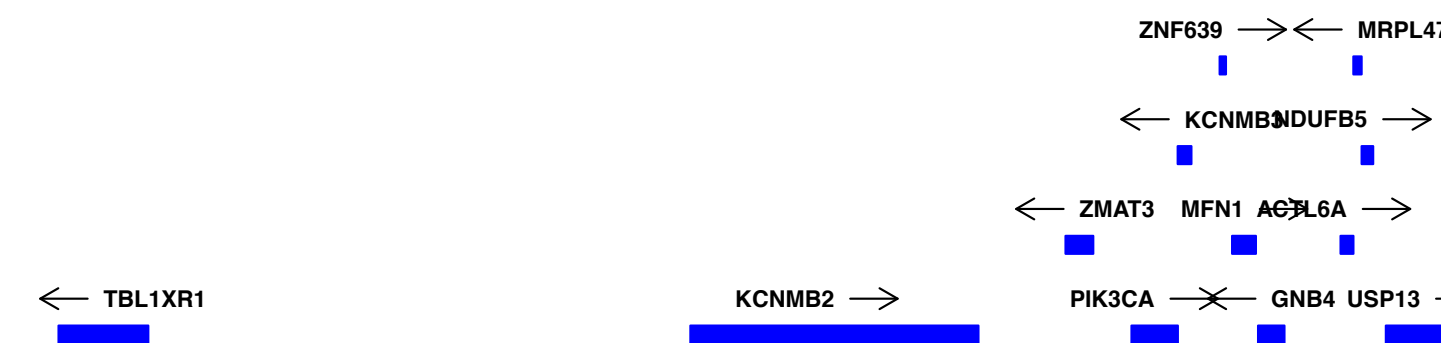
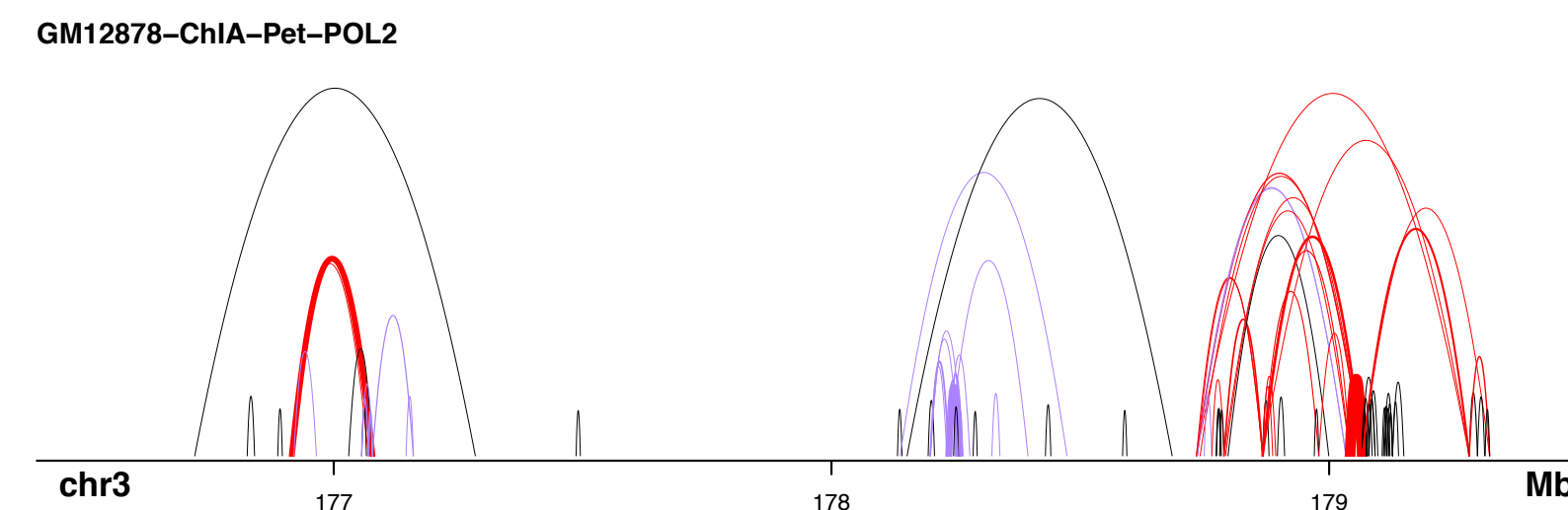
HiC



ChIA-PET: CTCF loops

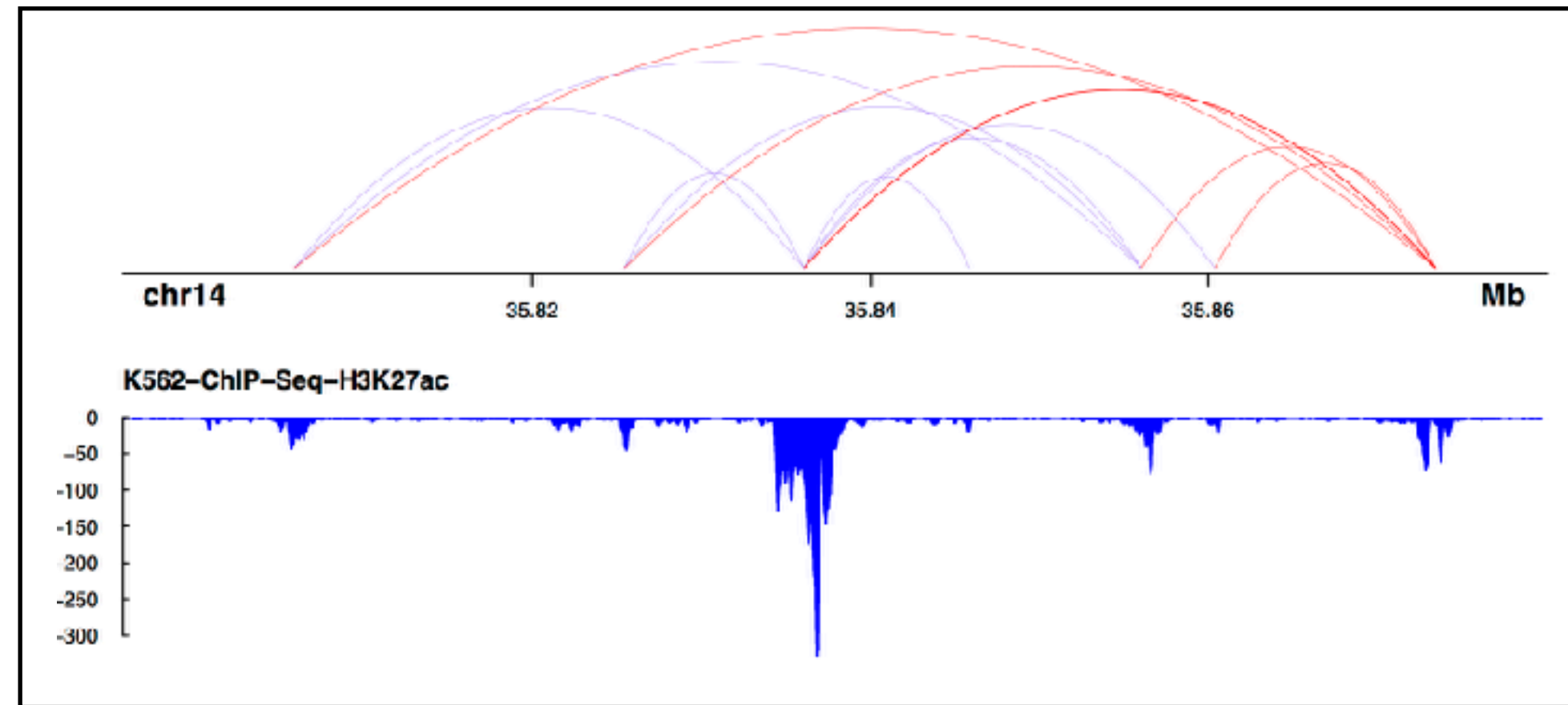


ChIA-PET: POL2 loops

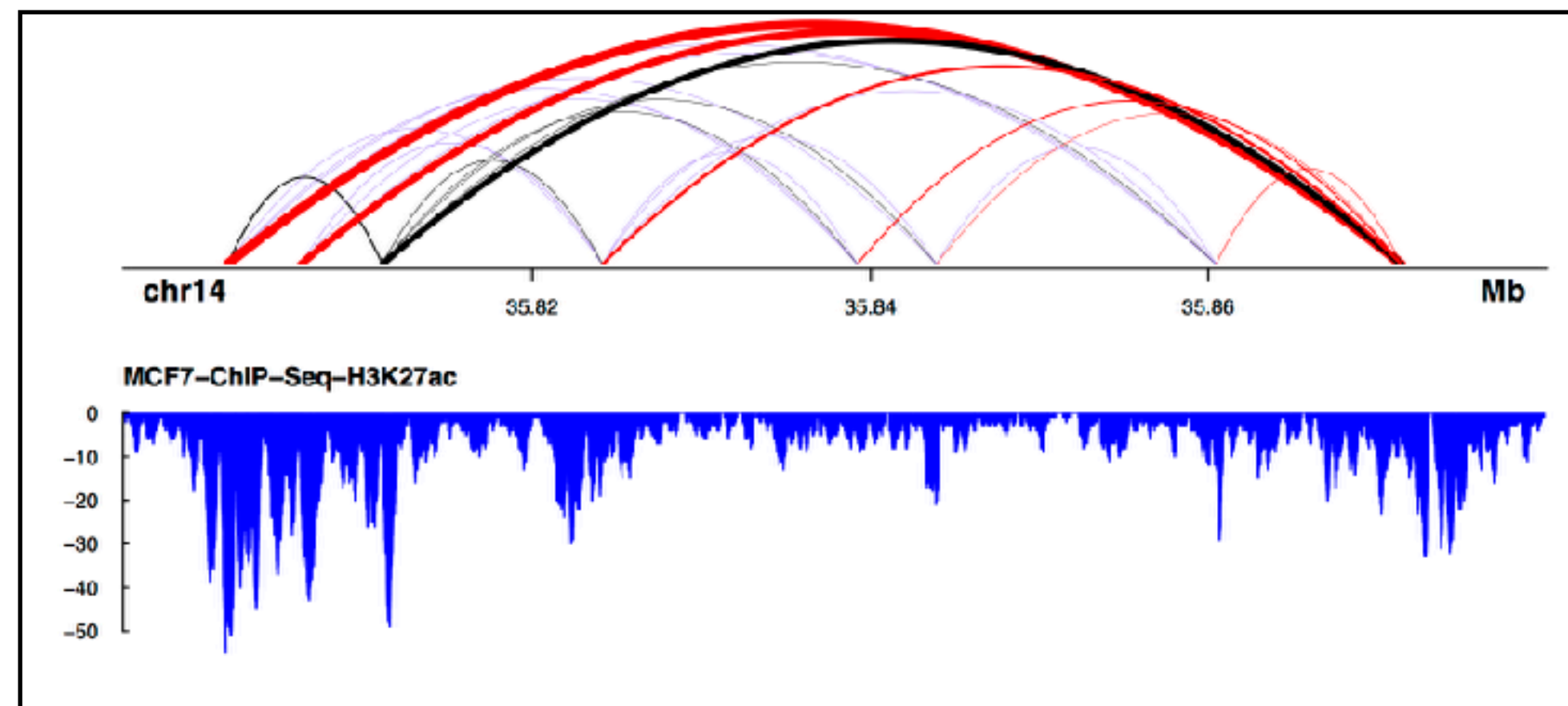


Differential topology analysis: POL2 ChIA-PET

K562



MCF7



← RP11-561B11.3 ← RP11-561B11.1 ← NFKB1A

H I III

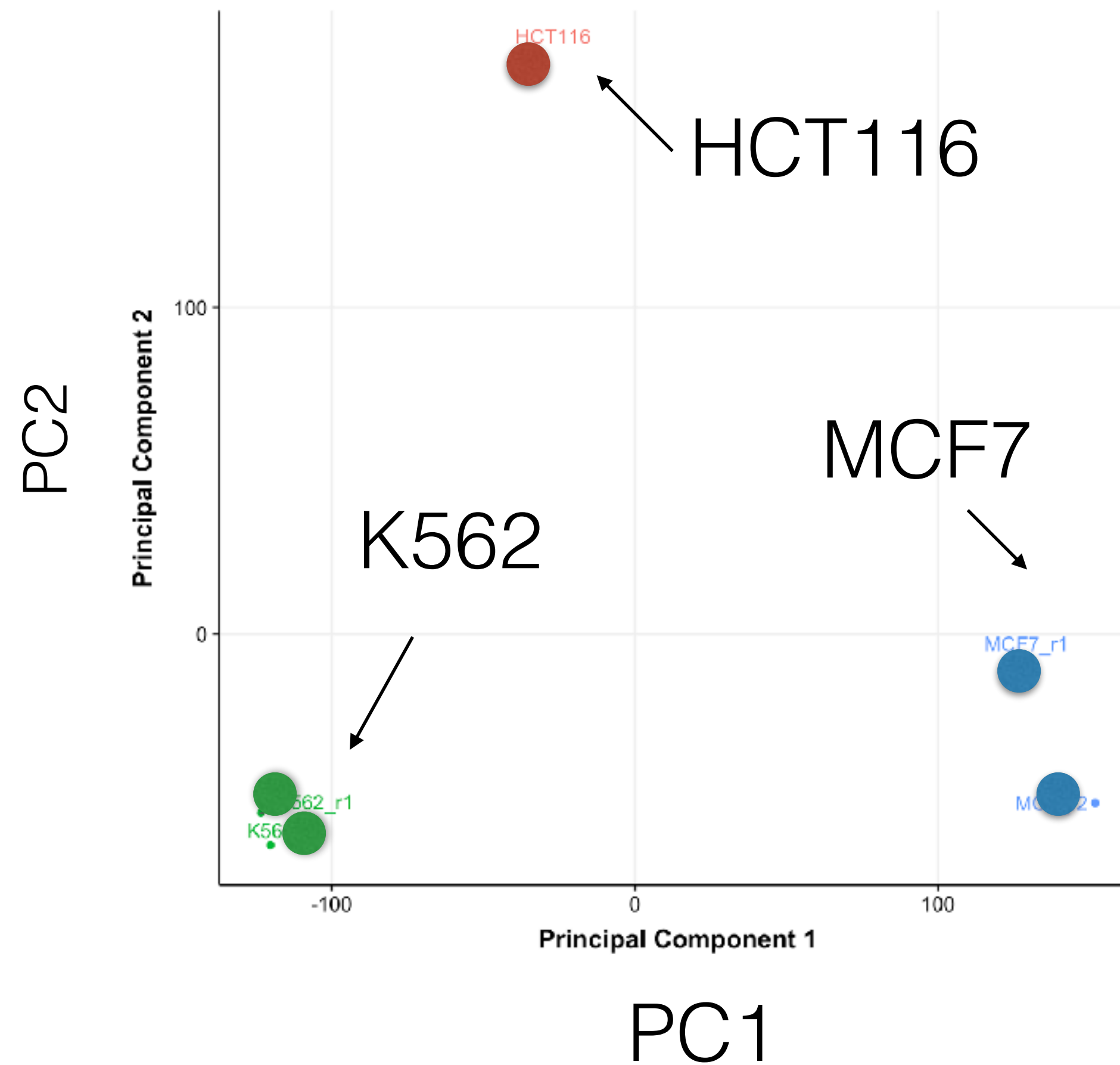
Legend

enhancer - promoter

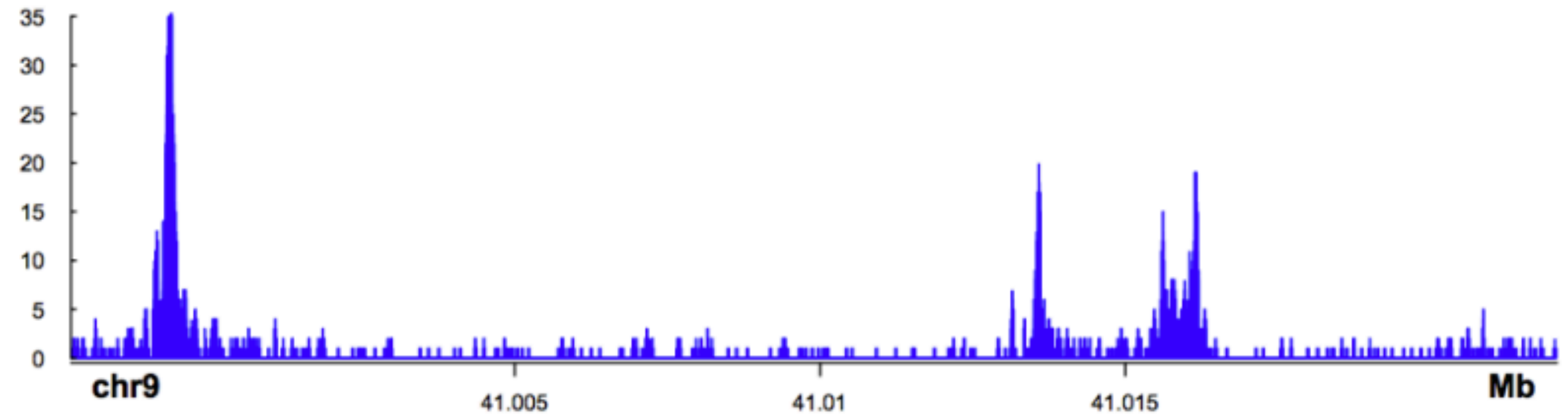
CTCF - CTCF

Pol2 ChIA-PET loops

PCA of ChIA-PET Counts Matrix



SMC1 ChIA-PET



Sample 1

PEAK

PEAK

PEAK

Sample 2

PEAK

PEAK

Sample 1

PETs



Sample 2

PETs



**Need to define a
common anchor set:**

<PEAK>

<PEAK>

<PEAK>

Differential loops: diffloop

	chr	start	end
[1]	1	10000	11000
[2]	1	12000	13000
[3]	1	17000	17600
[4]	1	21500	22000
	...		

Anchors

	left	right
[1]	1	2
[2]	1	3
[3]	2	3
[4]	2	4
	...	

Loops

	k1	k2	m1	m2	
[1]	4	3	1	0	Loop 1
[2]	1	2	6	2	Loop 2
[3]	2	0	7	8	...
[4]	0	1	1	0	
	...				

PET counts



Home

Install

Help

Home » Bioconductor 3.4 » Software Packages » diffloop

diffloop

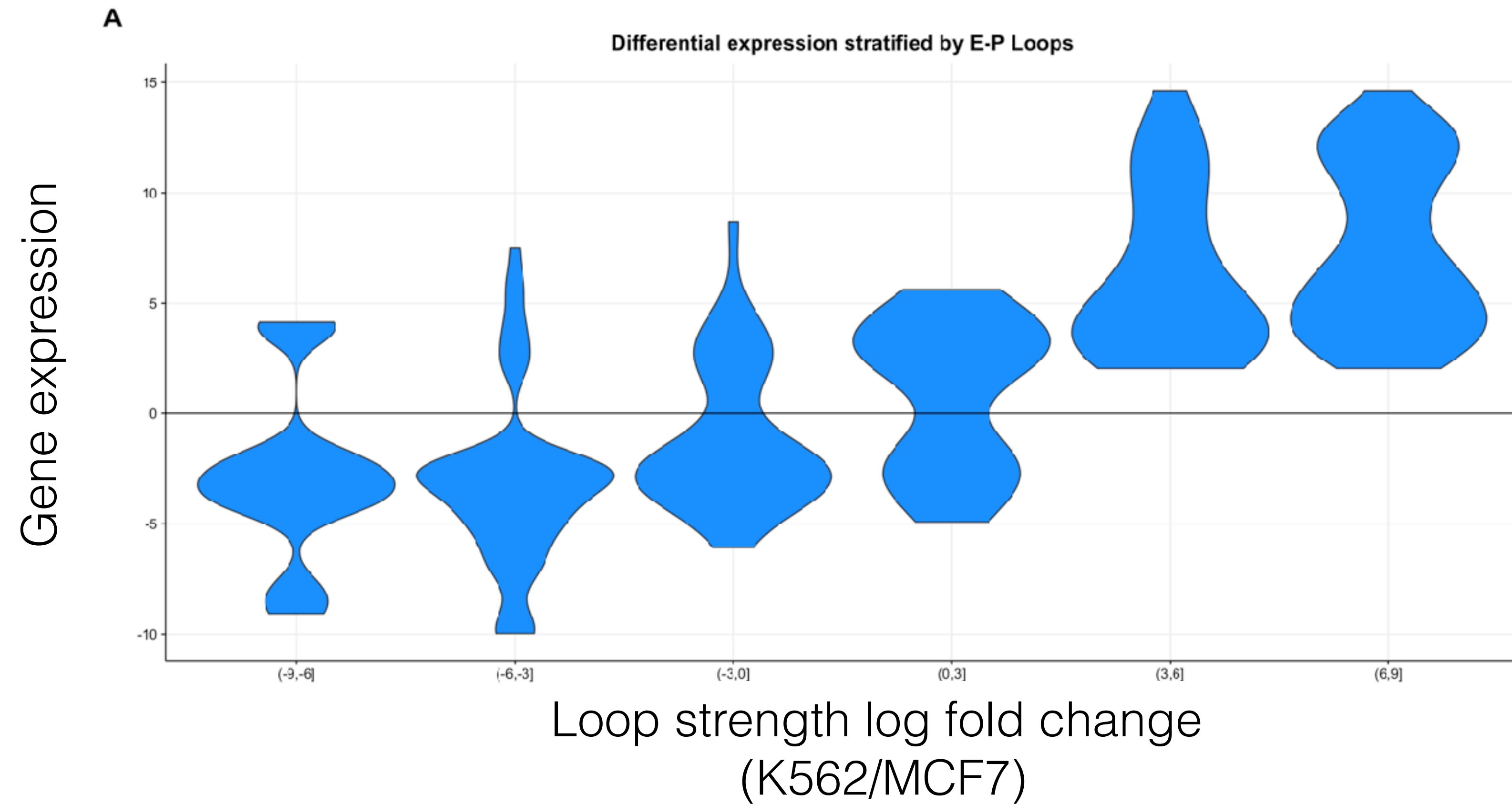
PET = paired end tag

<http://bioconductor.org/packages/diffloop/>

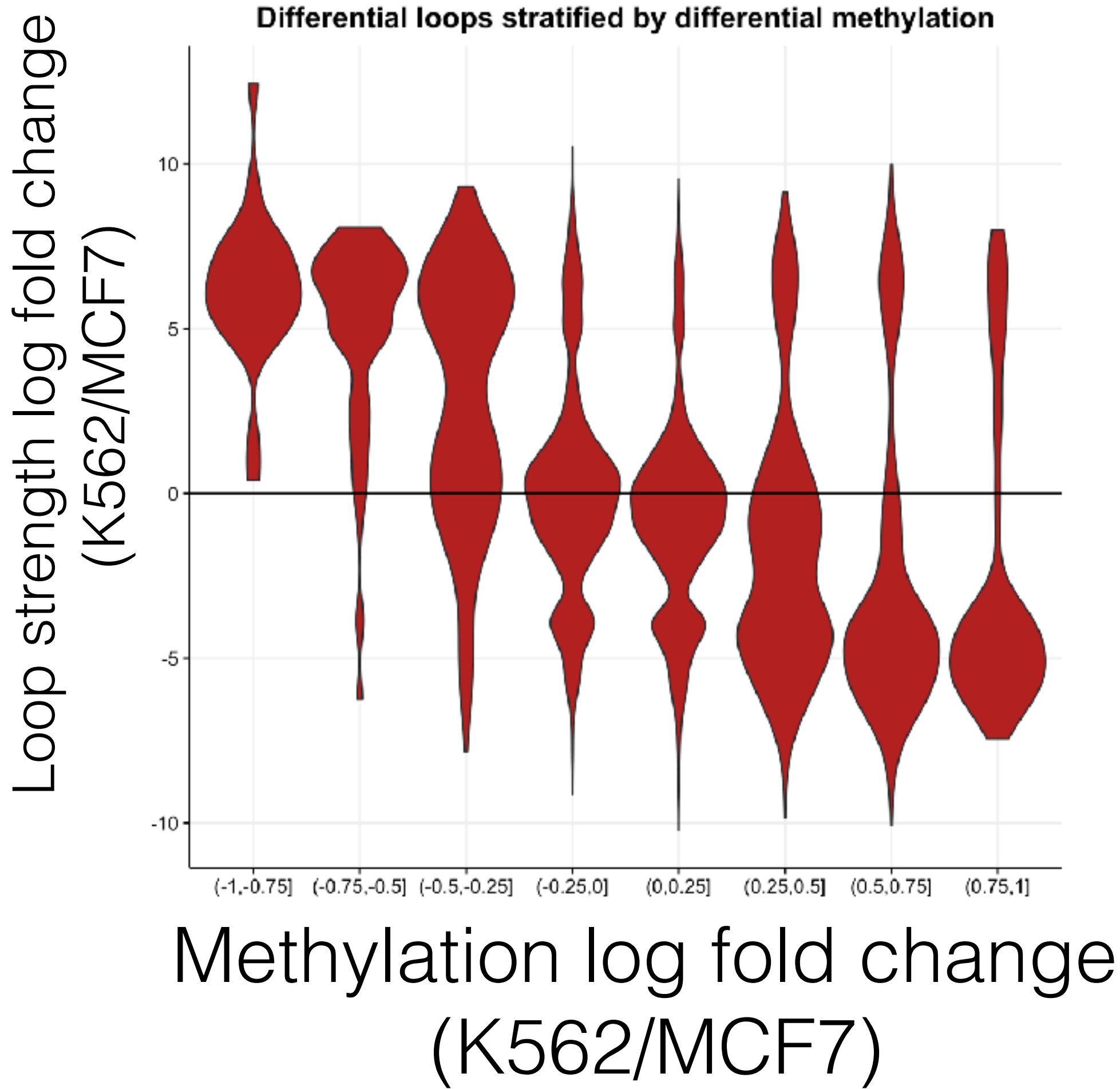
Genome-wide analysis of differential looping

- Genome-wide identification of **POL2** ChIA-PET loops in MCF7 and K562
- At an **FDR of 1%**, there are **> 2,600** differential loops
- Nearly **2,000** were classified as **enhancer-promoter loops** (another 500 p-p)
- Target genes in MCF7 enriched for **Estrogen Response Pathways**

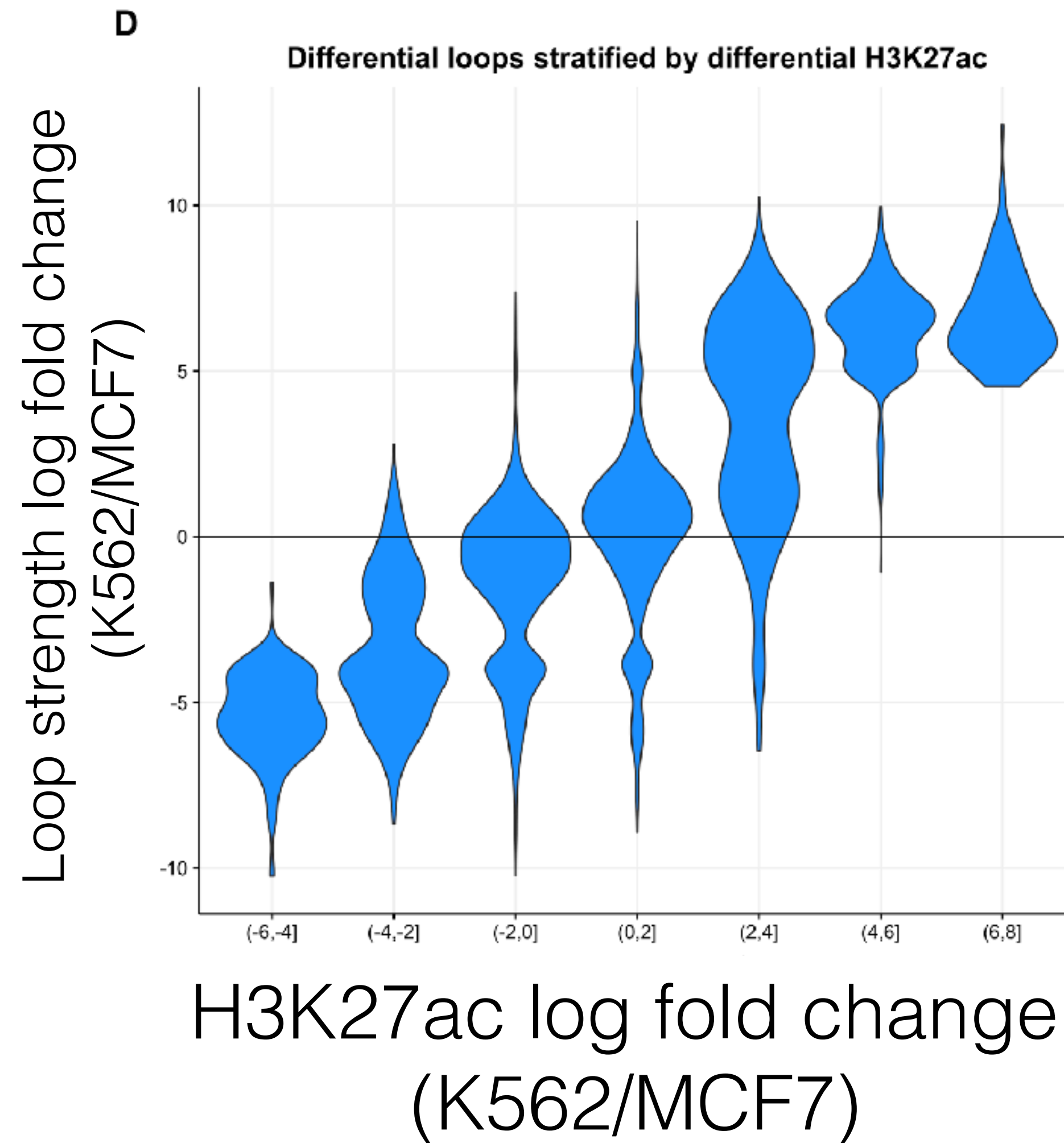
POL2 loop strength correlates with expression



Epigenetic correlates of enhancer-promoter looping: DNA Methylation



Epigenetic correlates of enhancer-promoter looping: H3K27Ac



HiChIP

NATURE METHODS | BRIEF COMMUNICATION



HiChIP: efficient and sensitive analysis of protein-directed genome architecture

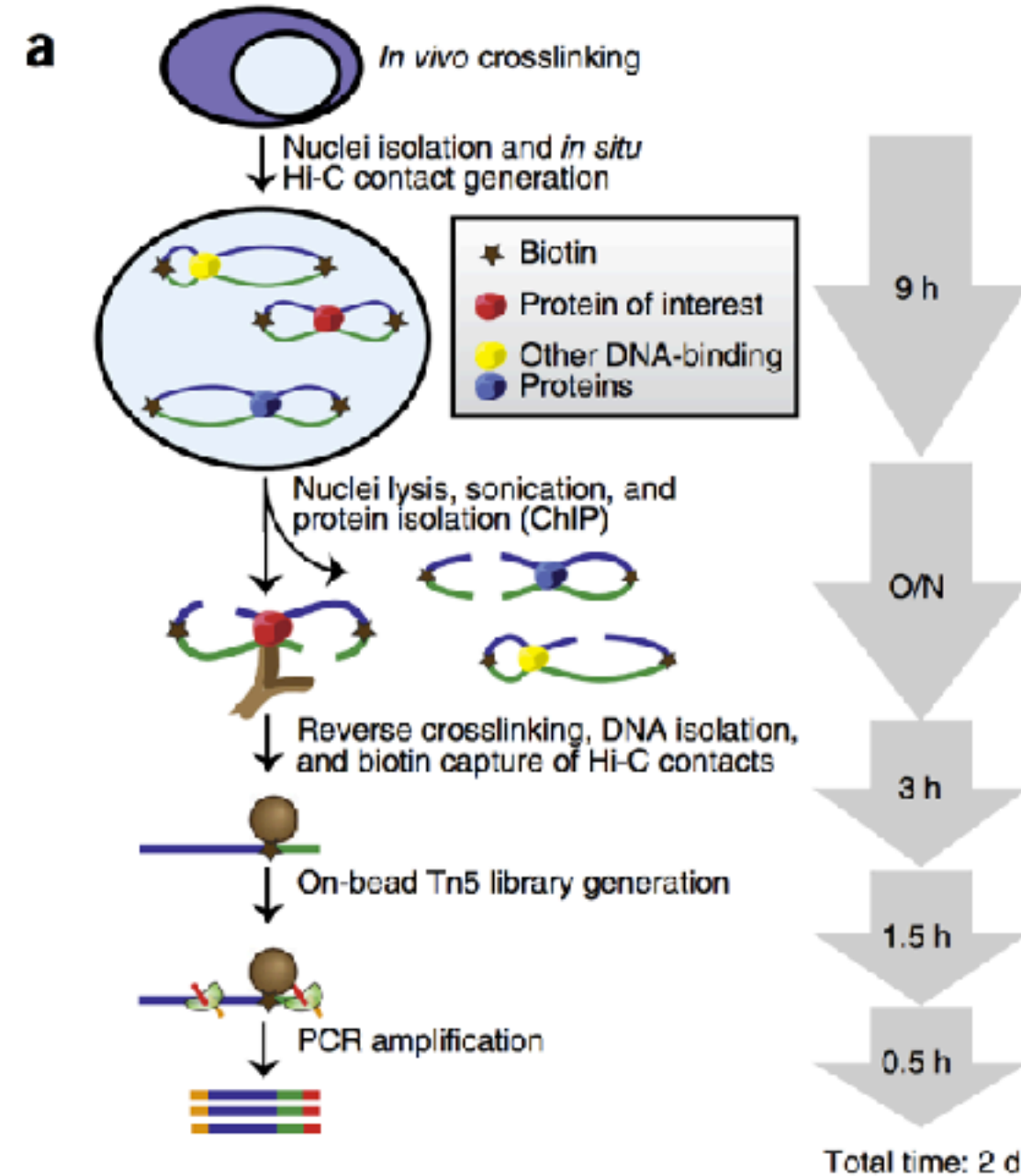
Maxwell R Mumbach, Adam J Rubin, Ryan A Flynn, Chao Dai, Paul A Khavari, William J Greenleaf & Howard Y Chang

[Affiliations](#) | [Contributions](#) | [Corresponding author](#)

Nature Methods **13**, 919–922 (2016) | doi:10.1038/nmeth.3999

Received 02 May 2016 | Accepted 10 August 2016 | Published online 19 September 2016

- Protocol allows for < 1 million cells
- Higher read efficiency than ChIA-PET
- Shorter protocol

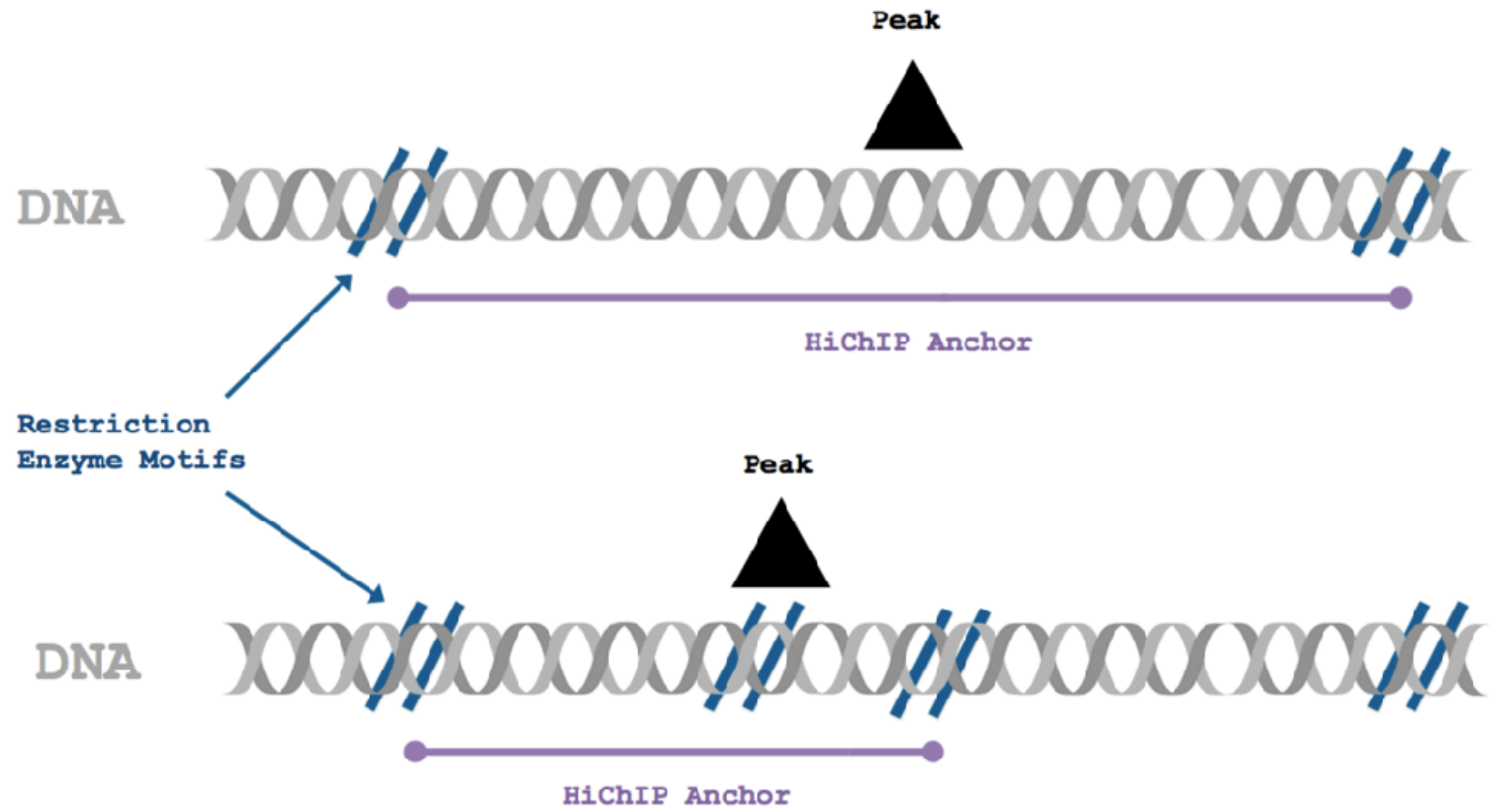


caleblareau added varying parameter comparison 18fb7fe on Dec 4, 2016
2 contributors

301 lines (234 sloc) 15.4 KB Raw Blame History

hichipper

This package is maintained by Caleb Lareau in the Aryee Lab. Source code is made freely available here and a packaged install version is provided through PyPi.



Acknowledgements

Caleb Lareau

Brad Bernstein

Rafael Irizarry

Zack McCaw

Sarah Johnstone

Alejandro Reyes

Jose Malagon Lopez

Esmat Hegazi